

(HUANJING KEXUE)

ENVIRONMENTAL SCIENCE

第39卷 第6期

Vol.39 No.6

2018

____ 中国科学院生态环境研究中心 主办

斜学出版社出版



ENVIRONMENTAL SCIENCE

第 39 卷 第 6 期 2018年6月15日

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西南喀斯特区植被恢复对土壤氮素转化通路的影响

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摘要: 氮素是生态系统重要的限制性养分元素之一. 研究氮素转化特征对于了解生态系统功能具有重要意义. 然而,目前对喀斯特地区氮素转化特征的认识十分有限. 同时,喀斯特地区正开展一系列的生态恢复工程措施,生态恢复将对土壤氮素转化过程产生何种影响尚不清楚. 为此,本研究在广西环江县喀斯特区域选取 3 种典型的植被恢复阶段(草地、灌丛、次生林),以农田为参照,采集 0~10 cm 深度的土壤样品,测定了土壤净氨化速率(净氨化率、真菌氨化和细菌氨化)、净氮矿化速率(净氮矿化、真菌矿化和细菌矿化)、净硝化速率(净硝化、自养硝化、异养硝化、真菌硝化、细菌硝化)及相关土壤理化指标,研究喀斯特区植被恢复对土壤氮素转化速率的影响. 结果表明,总体上喀斯特生态系统硝化速率很高,土壤无机氮主要以硝态氮形式为主,其中自养硝化和异养硝化分别占净硝化速率的80%和20%. 添加真菌和细菌抑制剂后,氨化速率增加,而硝化速率下降. 另外,随着植被的恢复,土壤氮矿化和硝化速率逐渐增加,而氨化速率逐渐下降. 其原因与不同植被恢复阶段的土壤有机碳、总氮、硝态氮、微生物量及氮获取酶的活性有密切关系. 这些发现为认识喀斯特生态系统氮素循环特征提供了关键的信息.

关键词:植被恢复; 氮素转化; 氮矿化; 自养硝化; 异养硝化; 喀斯特

中图分类号: X144 文献标识码: A 文章编号: 0250-3301(2018)06-2845-08 DOI: 10.13227/j. hjkx. 201710066

Effects of Vegetation Restoration on Soil Nitrogen Pathways in a Karst Region of Southwest China

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Abstract: Nitrogen (N) is an important element for plant growth in terrestrial ecosystems. Studying soil N cycling is crucial for understanding the structures and functions of an ecosystem. However, our knowledge of soil N dynamics in karst regions is still limited. In addition, while China's karst regions have conducted a series of vegetation restoration projects, the vegetation restoration effects on soil N pathways are still largely unknown. Therefore, this study selected four typical ecosystems representing four main vegetation restoration stages (i. e., cropland, grassland, shrubland, and forest) in a karst region in Huanjiang Province, southwest China. In these ecosystems, soil N pathways, including net ammonization rate (net ammonization, fungal ammonization, and bacterial ammonization), net nitrification rate (i. e., net nitrification, heterotrophic nitrification, autotrophic nitrification, fungal nitrification), and bacterial nitrification of the net nitrification rate was high in all ecosystems, but the ammonization rate was low, resulting in nitrite being the main inorganic N form in karst soil. Autotrophic and heterotrophic nitrification rates accounted for 80% and 20% of the net nitrification rate, respectively. After the addition of fungal and bacterial inhibitors, ammonization rates increased for all treatments, but the nitrification rates decreased. Following vegetation restoration, soil N mineralization and nitrification rates all increased, but the ammonization rates significantly decreased. This pattern was significantly correlated with soil organic carbon, total nitrogen, nitrate, microbial biomass, and the activity of N-acquisition enzymes in these ecosystems. Our findings provide very useful information for understanding soil N cycling in the karst regions.

Key words: vegetation restoration; nitrogen transformation; nitrogen mineralization; heterotrophic nitrification; autotrophic nitrification; karst

收稿日期: 2017-10-12; 修订日期: 2017-12-01

基金项目: 国家重点研发计划项目(2016YFC0502404); 国家自然科学基金项目(31400462, 31500405); 中国科学院百人计划项目 (Y523101030)

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氮是植物生长的必需元素, 也是陆地生态系统 最主要的限制性营养元素[1]. 由于生态系统碳、氮 循环之间的密切耦合关系,土壤氮素供应能力很大 程度上决定着全球变化背景下生态系统的固碳潜力 及退耕后植被演替的方向和进程[2].同时,土壤中 某些形态氮也是环境中重要的污染物质, 比如排放 到水体中的可溶性氮可导致水体富营养化[3],排放 到大气中的 NO, 可导致光化学烟雾[4]. 可见, 生态 系统氮状况的研究对于评估全球变化背景下生态系 统的固碳潜力、预测生态恢复方向与进程及污染防 控等一系列重大生态环境问题均具有重要意义. 土 壤有效氮的生产包括了一系列过程, 如氨化、硝化 和氮矿化过程等[5]. 这些过程是由一系列的真菌和 细菌参与的[6]. 以硝化作用为例, 铵态氮氧化为硝 态氮的过程是由自养和异养微生物共同驱动的. 相 比之下, 有机氮硝化为硝态氮的过程则主要是由异 养微生物驱动. 一般地, 自养硝化主要是由氨氧化 细菌(AOB)和硝氧化细菌(NOB)主导,而异养硝化 则是比较广泛的细菌和真菌主导[7]. 通过添加生物 抑制剂, 自养硝化、异养硝化、真菌硝化和细菌硝 化过程是可以被区分开的[8],而这样的区分对更好 地理解生态系统氮循环过程有重要的意义.

生态系统氮素转化过程受多种全球变化因素的 影响,土地利用方式改变是其中重要的影响因素之 一[9,10]. 目前关于土地利用方式改变后(如:退耕后 植被恢复)土壤氮素转化过程变化的研究已有不少 报道. 然而结果并不一致, Li 等[11]的研究发现净 氮矿化和硝化速率随演替而下降, 而氨化速率则不 断增加; Yan 等[12]的研究发现氮净矿化速率在演替 初期和后期高, 中期低, 而净硝化速率则呈现随演 替进程不断增加,且不同季节变化趋势差异明显. 生态系统类型、土壤母质和土壤类型是导致这些差 异的主要原因. 然而, 目前的研究主要集中在酸性 土壤区域,对中性偏碱性的喀斯特石灰土壤的氮素 转化特征的认识还十分不足. 此外, 对氮转化速率 的研究多数只关心净(或总)氮转化速率,对其具体 的通路, 如自养、异养硝化过程的研究缺乏深入 了解.

喀斯特是地表生态系统的重要组成部分,其中中国西南喀斯特区是世界最大、最集中连片的喀斯特区^[13].喀斯特山区独特的土壤条件(如高 pH、高钙与高缓冲能力、特殊水文条件等)决定了生态系统过程对于全球变化的响应有别于其他地区.如前期的研究表明,该地区的土壤氮含量很高,硝态氮

淋失严重,意味着该地区土壤氮饱和的可能性极大[14].同时,该地区正开展大规模的植被恢复工程,其带来的对土壤氮转化过程的影响也不甚清楚.为此,本文选取喀斯特地区4种不同土地利用方式的土壤(农田、草地、灌丛、次生林)为研究对象,通过对喀斯特地区不同植被恢复阶段的氮矿化、氨化、硝化、真菌矿化、细菌矿化、真菌氨化、细菌氨化、真菌硝化、细菌硝化、自养硝化和异养硝化进行测定,分析植被恢复过程对氮转化速率的影响及机制.本研究一方面可获取区域代表性数据,另一方面可揭示喀斯特山区土壤肥力的供应和保持机制,以期为喀斯特生态恢复实践提供理论支撑.

1 材料与方法

1.1 研究样地概况

研究区位于广西环江县(107°53′~108°05′E, 25°06′~25°12′N), 具有典型喀斯特峰丛洼地景观,属亚热带季风气候区. 年平均气温为 19.6~21.6℃,最低温出现在 1 月,约为 3.4~8.7℃,最高温出现在 7 月,约为 23.0~26.7℃. 年均降雨量为1 530~1 820 mm,分干湿季,4~8 月属于湿季,9 月至次年 3 月属于旱季. 研究区基岩主要为石灰岩和白云岩,土壤主要为石灰土[15].

1.2 土壤样品采集

土壤采集时间在 2014 年 7 月. 选择典型的农田、草地、灌丛和次生林这 4 种不同的植被恢复阶段. 其中农田种植的是玉米. 草地的恢复年限为 10 a 左右, 优势物种以五节芒(Miscanthus floridulus)、类芦(Neyraudia reynaudiana)等为主;灌丛封育年限为 15 ~ 20 a, 主要优势种为火棘(Pyracantha fortuneana)、金丝桃(Hypericum monogynum)和小果蔷薇(Rosa cymosa)等. 次生林封育年限为 30 a 以上, 主要的优势种有伞花木(Eurycorymbus cavaleriei)、樟树(Cinnamomum camphora)、厚壳桂(Cryptocarya chinensis)等.

每个植被恢复阶段建立 3 个 10 m×10 m 的样方,每个样方按 S 型布设 5 个样点,采集表层(0~10 cm)土壤样品.将 5 个样点采取的土壤样品混成一个混合土样,总共12 个土壤样品.新鲜土样放于冰盒中带回实验室,先用镊子挑去根和石块,过 2 mm 筛后分两部分保存.一部分保存在 4℃冰箱内用于土壤氮转化速率测定,另一部分经自然风干后用于土壤理化性质分析(表1).

衣I	个问性做恢复阶段下的工块偶性*	

Table 1 Soil properties in different vegetation restora	tion stag	tages	
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土壤属性	农田	草地	灌丛	次生林
SOC/g·kg ⁻¹	36. 07 ± 1. 72 ab	26. 47 ± 1. 79 b	$33.31 \pm 3.35 \text{ ab}$	40. 85 ± 5. 45 a
$TN/g \cdot kg^{-1}$	$2.08 \pm 0.38 \text{ b}$	$2.27 \pm 0.22 \text{ b}$	3.49 ± 0.53 a	$3.64 \pm 0.20 \text{ a}$
C/N	17. 79 ± 3.74 a	11.77 ± 1.81 a	$9.61 \pm 0.63 \text{ b}$	11. 20 ± 0.97 ab
K/cmol·kg ⁻¹	0.23 ± 0.07 a	0.19 ± 0.01 a	0.29 ± 0.08 a	0.21 ± 0.03 a
Ca/cmol·kg ⁻¹	$12.91 \pm 2.23 \text{ b}$	$18.60 \pm 2.26 \text{ a}$	18.39 ± 2.31 a	19.08 ± 4.38 a
Na/cmol·kg ⁻¹	0.31 ± 0.03 a	$0.34 \pm 0.04 \text{ a}$	0.38 ± 0.03 a	$0.33 \pm 0.02 \text{ a}$
Mg/cmol·kg ⁻¹	$2.80 \pm 1.77 \text{ b}$	$2.04 \pm 0.10 \text{ b}$	4.74 ± 2.08 ab	6. 27 ± 1.52 a
рН	6. 66 ± 0.30 ab	$6.92 \pm 0.11 \text{ a}$	$6.36 \pm 0.16 \text{ b}$	6.58 ± 0.08 ab
黏粒/%	13.53 ± 1.76 ab	$15.95 \pm 1.02 \text{ a}$	12. 62 ± 5.51 ab	$10.92 \pm 0.64 \text{ b}$
粉粒/%	77. 43 ± 3.52 a	$71.79 \pm 3.06 \text{ a}$	74. 46 ± 0.56 a	73. 99 ± 1. 68 a
砂粒/%	$9.04 \pm 5.22 \text{ a}$	12. 26 ± 2.07 a	12. 92 ± 5.00 a	15. 09 \pm 2. 16 a
微生物 C/mg·kg -1	$288 \pm 8 \text{ c}$	$293 \pm 33~\mathrm{bc}$	$365 \pm 17 \mathrm{b}$	$836 \pm 51 \text{ a}$
微生物 N/mg·kg - 1	$38 \pm 0.5 c$	$58 \pm 10 \text{ bc}$	$62 \pm 8 \text{ b}$	198 ± 16 a
BG/nmol·(g·h) -1	$29.5 \pm 1.4c$	$43.6 \pm 2.6 \text{ b}$	$49.9 \pm 5.8b$	86. 1 ± 11. 8a
CBH/nmol·(g·h) -1	$4.3 \pm 0.7c$	$6.2 \pm 0.6c$	$10.2 \pm 2.5 $ b	15. 2 ± 2. 1a
LAP/nmol·(g·h) -1	$0.8 \pm 0.1c$	$1.1 \pm 0.3c$	$2.7 \pm 0.6b$	$3.4 \pm 0.9a$
NAG/nmol·(g·h) -1	$11.2 \pm 1.5 c$	$30.~3\pm3.~6\mathrm{bc}$	$26.4 \pm 3.1b$	$40.8 \pm 6.3a$
Urease/nmol·(g·h) -1	135. $4 \pm 8.0 ab$	$126.9 \pm 44.6b$	$183.7 \pm 12.0a$	$268.0 \pm 64.3a$

¹⁾表中数值为平均值 ± 标准差;不同字母表示差异性显著;BG:β-葡糖苷酶;CBH:α-纤维素酶;LAP:亮氨酸氨基肽酶;NAG:β-乙酰葡糖胺糖苷酶;Urease;脲酶

1.3 实验方法

1.3.1 土壤潜在净氮转化速率测定

土壤潜在净氮转化速率的测定参考文献[16]的方法. 具体步骤如下:将新鲜土样分为 ABCDE 5 组. A 组:称 5 g 土测定土壤 NH⁺ 和 NO⁻。浓度作为培养的初始值. B 组:将 10 g 土壤放入 250 mL 培养瓶中,不添加任何物质; C 组:将 10 g 土壤放入 250 mL 培养瓶中,添加纯度为 99%的乙炔. 乙炔的注入量为 2% (体积比),用以抑制微生物的自养硝化; D 组:将 10 g 土壤放入 250 mL 培养瓶中,添加 2 mL 的放线菌酮试剂,用以抑制土壤的真菌活动; E 组:将 10 g 土壤放入 250 mL 培养瓶中,添加 2 mL 的链霉素试剂,用以抑制土壤的细菌活动.

所有土壤样品,调节含水量至土壤最大持水率的 85%. 用聚乙烯膜将 B-E 组的瓶口包扎. 除 B 组的外,C-E 组在膜上扎 5 个小孔以保证透气性,然后将 B-E 组所有土壤置于 25℃培养室中培养 7 d. 培养期间,每隔两天调节一次含水率. 培养结束后,加入 2 $\operatorname{mol·L}^{-1}$ KCl 溶液 100 mL ,于温度为 25℃的恒温振荡机内振荡 1 h 后进行过滤,滤液用 AA3 连续流动分析仪(Fiastar 5000) 测定土壤中的 NH_4^+ 和 NO_3^- 含量. 土壤潜在净氮转化速率[以 NH_4^+ NO_3^- 含量. 土壤潜在净氮转化速率[以 NH_4^+ NO_3^- 含量. 土壤潜在净氮转化速率[以 NH_4^+ NO_3^- 含量.

净氮矿化速率 =
$$[(B \text{ 组的 } NH_4^+ + NO_3^-) - (A \text{ 组的 } NH_4^+ + NO_3^-)]/7 \text{ d}$$

净氨化率 =
$$\frac{[(B \text{ 组的 } NH_4^+) - (A \text{ 组 } NH_4^+)]}{7 \text{ d}}$$

净硝化率 = $\frac{[(B \text{ 4l } NO_3^-) - (A \text{ 4l } NO_3^-)]}{7 \text{ d}}$

自养硝化速率 = [(C组的NO; 浓度)-(A组的NO; 浓度)]/7 d

异养硝化速率 = 净硝化速 - 自养硝化速率 细菌异养硝化速率 = $[(D \text{ 组的 NO}_3^- \text{ 浓度}) - (A \text{ 组的 NO}_3^- \text{ 浓度})]/7 d$

真菌异养硝化速率 = $[(E ext{ 组的 NO}_3^- ext{ 浓度})$ -

(A组的NO; 浓度)]/7 d

细菌氨化速率 = [(D组的NH₄ 浓度)-

(A组的NH₄ 浓度)]/7 d

真菌氨化速率 = [(E组的 NH₄ 浓度) -

(A组的NH⁺浓度)]/7 d

1.3.2 土壤的理化指标分析

土壤理化指标的测定参考刘光崧等的方法^[18]. 土壤 pH 采用 pH 计测定(土水比为1:2.5);土壤有机碳(SOC)采用重铬酸钾氧化-外加热法(油浴)测定;全氮(TN)采用半微量开氏法-流动注射仪(FIAstar 5000)测定;交换性 K⁺、Ca²⁺、Na⁺、Mg²⁺采用乙酸铵交换-原子吸收分光光度法测定.土壤机械组成采用激光衍射粒度分析仪(Mastersizer, 2000, Malvern, UK) 测定. 土壤微生物量 C 和 N (MBC 和 MBN)采用氯仿熏蒸法测定. 此外还测定了与土壤有机氮分解相关的 5 种酶的活性. 这些酶包括 β -葡糖苷酶 (BG)、 α -纤维素酶 (CBH)、亮氨酸氨基肽酶 (LAP)、 β -乙酰葡糖胺糖苷酶 (NAG)和脲酶 (Urease). 这些酶的测定方法和功能见文献 [19~21].

1.4 数据处理与分析

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利用单因素方差分析(One-way ANOVA)检验不同土地利用方式下土壤氮转化速率的差异,并采用 LSD 方法进行显著性多重比较,差异显著性水平

为 *P* < 0.05. 利用线性回归分析土壤氮转化速率与土壤理化指标之间的相关性. 以上分析在 SPSS 19.0 软件中进行.

2 结果与分析

在4个喀斯特生态系统中,土壤无机氮均以硝态氮为主,占到总无机氮的(65%~93%)(表2,A组).土壤培养7d后,硝态氮在所有处理中均升高,其中B组的升高幅度最大;C组、D组和E组相比B组有明显的下降.铵态氮在所有处理中也均显著升高,其中C组、D组和E组的升高幅度大于B组.

表 2 不同处理下土壤硝态氮和氨态氮浓度的差异¹⁾/mg·kg⁻¹

Table 2 Con	centrations of	soil	nitrate	and	ammonia	under	different	treatments/	mg•k	g -	
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处理	A 组((0 d)	B 组((7 d)	C 组	(7 d)	D组	(7 d)	E 组	(7 d)
处理	硝氮	氨氮	硝氮	氨氮	硝氮	氨氮	硝氮	氨氮	硝氮	氨氮
农田	9.5(0.9)	2.2(1.0)					34.69(2.1)			
草地	2.8(0.4)	1.5(0.2)	16.2(1.9)	6.0(0.5)	5.0(0.9)	9.0(0.5)	11.82(1.8)	27.7(5.6)	10.02(0.7)	36.4(3.4)
灌丛	18.2(0.6)		75.3(1.7)							
次生林	36.4(3.0)	2.5(0.4)	188.8(23.3)	1.0(0.2)	71.7(9.3)	3.7(1.6)	94. 59 (15. 2)	3.4(0.5)	76.63(8.6)	5.8(1.5)

1)括号内数字表示平均值,括号外表示标准差

2.1 土壤的净氨化、细菌氨化与真菌氨化速率

4种喀斯特生态系统类型的土壤净氨化速率均较低,介于 -0.16 ~0.63 mg·(kg·d) -1(图1).方差分析结果表明,随着植被的恢复,土壤净氨化速率显著降低,甚至在次生林阶段表现为负值(图1).通过添加细菌和真菌抑制剂计算出来的细菌氨化[0.37 ~ 3.88 mg·(kg·d) -1] 与真菌氨化速率[0.11 ~ 3.37 mg·(kg·d) -1] 均高于总的净氨化速率(图1).细菌氨化速率在次生林阶段显著低于农田、草地和灌丛,而后3个阶段之间没有显著差异.真菌氨化速率表现出和细菌氨化速率一致的规律(图1).

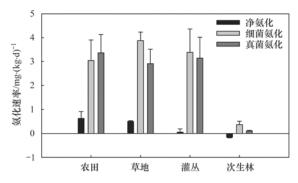


图 1 不同土地利用方式对土壤的净氨化、 细菌与真菌氨化速率的影响

Fig. 1 Effect of different land use types on the net rates of ammonification, fungal and bacterial ammonification

2.2 土壤的总硝化、自养与异养硝化、真菌硝化和细菌硝化速率

4 种喀斯特生态系统类型的土壤净硝化速率均较高,介于 1.49~16.94 $mg \cdot (kg \cdot d)^{-1}$,其中自养硝化[1.24~13.02 $mg \cdot (kg \cdot d)^{-1}$]相比异养硝化[0.25~3.92 $mg \cdot (kg \cdot d)^{-1}$]占到总净氨化速率比重的 76.8%~83.3%(图 2).细菌硝化介于 0.80~4.46 $mg \cdot (kg \cdot d)^{-1}$ 之间,平均约占净氨化速率的41%;真菌硝化速率介于 1~6.46 $mg \cdot (kg \cdot d)^{-1}$ 之间,平均约占 59%(图 3).以上几种硝化速率类型均表现为随着植被恢复不断增加的趋势,其中在草地最低,次生林最高.

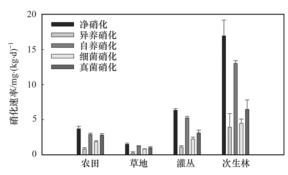


图 2 不同土地利用方式对土壤的总硝化、 自养与异养硝化和真菌与细菌硝化的影响

Fig. 2 Effects of different land-use types on the net rates of soil nitrification, autotrophic and heterotrophic nitrification and fungal and bacterial nitrification

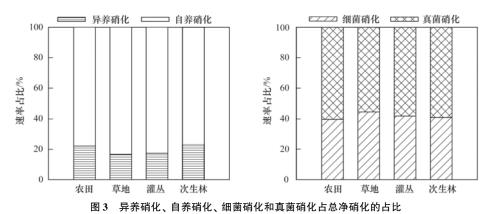
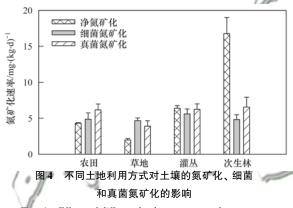


Fig. 3 Contributions of heterotrophic, autotrophic, bacterial, and fungal nitrification to the total net nitrification

2.3 土壤的总氮矿化、真菌氮矿化与细菌氮矿化 速率

土壤净氮矿化速率介于 $2.00 \sim 16.78$ mg·(kg·d) $^{-1}$, 在次生林阶段显著低于农田、草地和灌丛(图 4). 细菌氮矿化 $[0.37 \sim 3.88$ mg·(kg·d) $^{-1}$] 与真菌氮矿化速率 $[0.11 \sim 3.37$ mg·(kg·d) $^{-1}$] 也表现出相同的规律. 而且除了次



g. 4 Effects of different land-use types on the net rates of soil nitrogen-mineralization, bacterial and fungal nitrogen-mineralization

生林外,真菌和细菌氮矿化均高于总的净氮矿化 速率

2.4 土壤氮转化速率与土壤属性的关系

相关性分析的结果显示, 土壤氮转化速率与土 壤有机碳、总氮、硝态氮、微生物量C、微生物量N 及5种与氮获取相关的酶的活性关系密切,而与土 壤pH、碳氮比、铵态氮、土壤钾钙钠镁的含量无显 著相关(表3). 其中, 土壤有机碳与净氮矿化、真 菌矿化、细菌矿化、净硝化、异养硝化、自养硝化、 真菌硝化和细菌硝化呈显著正相关; 土壤总氮与净 氮矿化、真菌矿化、细菌矿化、净氨化、净硝化、异 养硝化、自养硝化、真菌硝化和细菌硝化呈显著正 相关;而土壤硝态氮与测定的所有氮转化速率均成 正相关. 微生物量 C 与净氮矿化、细菌矿化、净氨 化、细菌氨化、净硝化、自养硝化成正相关. 微生 物量 N 与净氮矿化、净氨化成正相关. 5 种酶中, LAP 的相关性最强,与净氮矿化、真菌矿化、细菌 矿化、细菌氨化、净硝化、异养硝化、自养硝化和 细菌硝化均呈显著正相关.

表 3 土壤氮转化速率与土壤属性的相关分析结果1)(P值)

	Table 3	Results of	correlation a	analysis for	the relation	ships betwe	en soil nitro	ogen transfo	rmation rate	and soil p	roperties	
	pН	SOC	TN	NO_3^-	$\mathrm{NH_4}^+$	MBC	MBN	BG	CBH	LAP	NAG	Urease
净氮矿化	0. 761	0.001	0.001	0.000	0.752	0.007	0. 027	0. 018	0.001	0. 005	0. 073	0.002
真菌矿化	0. 488	0.001	0.002	0.000	0.315	0. 247	0.308	0. 279	0.058	0. 047	0.350	0. 171
细菌矿化	0. 547	0.002	0.001	0.000	0. 199	0.025	0. 143	0. 019	0.018	0.001	0.014	0.059
净氨化	0. 536	0. 128	0.032	0.004	0.556	0.007	0.016	0.009	0.040	0. 192	0. 014	0.031
真菌氨化	0. 431	0.060	0.062	0.008	0.459	0.394	0.342	0. 379	0. 126	0. 157	0. 241	0. 501
细菌氨化	0. 966	0.078	0. 128	0.005	0. 441	0.049	0. 200	0. 029	0.077	0.014	0.009	0. 161
净硝化	0. 741	0.001	0.001	0.000	0.734	0.047	0. 105	0.087	0.010	0.012	0. 221	0.015
异养硝化	0.742	0.001	0.002	0.000	0.727	0. 135	0. 189	0. 218	0.053	0.025	0. 383	0.062
自养硝化	0.748	0.003	0.001	0.000	0.773	0.038	0.096	0.072	0.007	0.011	0. 198	0.011
真菌硝化	0.719	0.000	0.001	0.000	0. 532	0.315	0. 507	0. 395	0.160	0. 182	0.769	0. 111
细菌硝化	0.647	0.002	0.003	0.000	0. 540	0.336	0. 468	0.425	0. 127	0.033	0.708	0. 176

1)黑体数据为显著相关(P<0.05); 氮转化速率与土壤钾钙钠镁的含量、土壤质地无显著相关, 故分析结果没放在表内

3 讨论

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3.1 喀斯特地区土壤氮转化的特征

喀斯特钙质土壤的无机氮主要以硝态氮形式存在,而铵态氮所占比例很低(表2). 该结果与其他钙质土壤地区的研究一致^[22],而与酸性土壤的研究相反:一般酸性土壤中的无机氮以铵态氮为主^[23]. 偏碱性的钙质土壤硝态氮含量高的原因可能是较高的硝化速率导致的^[24]. 一般认为,硝化作用在 pH <5 的时候很低,而随着土壤 pH 值的升高(从4升至8)硝化作用不断增加^[25]. 以此相符,本研究中喀斯特钙质土壤的硝化速率很高,相反地,氨化速率非常低,甚至在次生林中表现出负值. 较高的硝化速率使得氨化产生的铵态氮很快被消耗. 当总硝化速率超过了总氨化速率时,净氨化速率则会表现为负值.

研究发现在添加了真菌或细菌抑制剂后,无论是真菌还是细菌氨化都比总的净氨化速率高.这和一般的认知相反.一般地添加抑制剂后,氨化速率都会下降.比如,Trap等[16]的研究发现,在添加抑制剂后,真菌氨化和细菌氨化均比净氨化速率低.导致本研究中真菌氨化和细菌氨化均高于对照的净氨化速率的原因还不清楚.推测可能是由于添加生物抑菌剂在杀死硝化细菌的同时也杀死了氨化细菌.由于喀斯特地区的硝化速率较高,相同比例的硝化细菌被杀死后,可能导致硝化速率下降,使得铵态氮累积,最终表现为净氨化速率的上升.本研究确实发现添加真菌和细菌抑制剂后铵态氮增加了,但硝化速率和硝态氮浓度均下降(表2),这个结果支持了上述的解释.

本研究还发现自养硝化相比异养硝化占更重要的地位.这和目前的认知是一致的.自养硝化细菌在低 pH 的环境中含量很低,所以一般认为在酸性土壤中自养硝化很弱,相反地,在中性或碱性土壤(pH 为 7~8),自养硝化速率达到最高^[4].此外,在喀斯特生态系统中细菌和真菌对硝化的贡献均较高,分别占41%和59%.之前的研究结果表明,喀斯特地区土壤的细菌约占总微生物量的11.3%~63.6%^[26],略低于真菌的比例.这部分支持了以上的结果.然而,进一步的研究有待揭示起作用的真菌与细菌的具体种类及其作用机制.

3.2 植被恢复对喀斯特土壤氮素转化速率的影响 随着植被恢复,土壤净氮矿化速率(包括其子 通路)和净硝化速率(包括其子通路)从农田到森林

不断增加, 而净氨化作用(包括其子通路)不断下 降. 氮矿化速率和硝化速率随着植被恢复不断上升 的规律与之前的研究一致. 比如, 刘欣等的研究发 现[27],随着植被的正向演替(草丛→灌丛→次生林 →原生林),土壤氮净矿化速率、净硝化速率均呈 增加趋势. 然而, 其测定的净氨化速率也呈现增加 的趋势, 这与本研究的结果相反. 目前关于喀斯特 地区土壤氮素转化特征随植被恢复变化的研究较 少. 因此, 到底氨化速率随植被恢复后如何改变还 需进一步的研究. 本研究推测净氨化作用下降可能 是由于总硝化作用的上升幅度高过了总氨化作用, 从而导致氨氮的生成赶不上其消耗,最终表现为净 氨化作用的下降. 虽然本研究并没有测定总氨化和 总硝化速率, 但是相关分析显示, 土壤净氨化与土 壤硝态氮成显著负相关关系. 这个结果部分支持了 上述解释.

而导致净氮矿化和硝化速率上升的原因可能有 两个. 第一个原因与土壤碳氮水平有关. 一般认为 土壤中的有机碳和氮的增加会导致总氮矿化或者硝 化的作用增加, 因为其增加了微生物的能量和底物 来源[27]. 本研究中土壤有机碳,有机碳与净矿化硝 化有显著的正相关关系,说明土壤有机碳、氮含量 的增加确实对土壤净矿化消化速率的上升有显著贡 献. 与本研究的结果一致, 刘欣等[27] 及周练川 等[28] 也发现相似的结果. 第二, 微生物量及酶活性 也是导致土壤氮矿化和硝化速率上升的关键因素. 微生物是土壤生物化学循环的主要驱动者[29],因 此土壤微生物量的大小直接决定了土壤氮素转化的 速率. 本研究发现土壤氮矿化和硝化速率与土壤微 生物量 C 有显著的正相关关系, 支持了上述观点. 同时,土壤有机氮的释放是在一系列氮获取酶的催 化下进行的[20]. 这些酶对土壤有机氮的降解被认 为是土壤氮循环的主要限速步骤[20]. 通过测定与 土壤有机氮分解相关的酶的活性(包括 BG、CBH、 LAP、NAG、Urease), 发现这些酶的活性均与土壤 氮转化速率有极强的相关性,尤其是蛋白质水解酶 (LAP酶)相关性最高(表3),说明蛋白质的降解可 能是土壤氮矿化的最重要的限速步骤.

4 结论

- (1)喀斯特区土壤硝化作用强烈,导致土壤无机氮主要以硝态氮为主.
- (2) 喀斯特土壤硝化作用主要以自养硝化为主,真菌和细菌均对土壤硝化有重要贡献.

(3)随着植被恢复,土壤净氮矿化速率(包括 其子通路)和净硝化速率(包括其子通路)从农田到 森林不断增加,而净氨化作用(包括其子通路)不断 下降.

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HUANJING KEXUE

Environmental Science (monthly)

Vol. 39 No. 6 Jun 15, 2018

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