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螺-草水质净化系统氮素环境归趋的实验研究

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摘要:通过构建螺-草模拟系统并利用稳定同位素示踪技术研究池塘螺-草水质净化系统中氮素的环境归趋,结果表明,以底泥为基质的螺-草系统中,实验结束后苦草湿重增加了580%,分株数增加了6.6 株,苦草根部吸收储存了1.07%的¹⁵N,苦草茎叶吸收储存了7.74%的¹⁵N,环棱螺吸收储存较少,只占0.06%,底泥滞留了5.73%的¹⁵N.结果分析表明:螺-草水质净化系统中苦草对水体中溶解态氮的吸收较少,沉积物是苦草生长的主要营养源;水体中氨氮主要通过沉积物-水界面进行迁移转化,大部分被苦草根系吸收利用转化为生物体,少部分通过硝化/反硝化作用去除,其余则滞留于沉积物;苦草是系统中氮素去除的最终载体,环棱螺的存在通过促进苦草生长及加强泥-水界面硝化和反硝化作用来加快系统中氮素的去除.因此,在养殖的不同阶段合理配置螺-草结构是整个养殖过程中水质调控的关键.

关键词:环棱螺; 苦草; 营养盐; 环境归趋; 稳定同位素示踪

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Experimental Study on the Environmental Fate of Nitrogen in Snail-Macrophyte Ecosystem for Water Purification

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Abstract: A snail-macrophyte simulation system was built and isotope tracer technique was adopted to study the environmental fate of nitrogen in snail-macrophyte purification system, the results showed that: Vallisneria spiralis increased its wet weight by 580% and its number by 6.6 ramets, moreover, Vallisneria spiralis absorbed 1.07% ¹⁵N by the roots and 7.74% by stems and leaves, while Bellamya only absorbed 0.06%. And 5.73% ¹⁵N was retained in the sediment. Through analyzing of the results, it indicated that: in such simulation system, sediment was the main nutrition source for the growth of Vallisneria spiralis, which absorbed only few dissolved nitrogen from water; ammonium nitrogen in water was transformed mainly in the sediment-water interface, most of which was absorbed by Vallisneria spiralis, a small amount was removed through nitrification and denitrification, and the rest was kept by sediment; Vallisneria spiralis was final vector for removing nitrogen in the system, and Bellamya could also boost the growth of Vallisneria spiralis and strengthen the processes of nitrification and denitrification, thus promoting the nitrogen removal from the system indirectly. So, during the period of culture, rational allocation of snail-macrophyte structure in different stages plays an important role in controlling water quality in ponds.

Key words: Bellamya; Vallisneria spiralis; nutrition; environmental fate; stable isotopic tracing

池塘河蟹养殖,为调控水质一般在池塘中种植一定面积的沉水植物并配养一定量的大型底栖动物如螺类等. 螺类与沉水植物既是河蟹的天然饵料更是重要的水质调控生物^[1,2],研究表明:螺类在底层的摄食、掘穴及爬动行为促进了底泥中有机碎屑的分解,加速了泥水界面的物质交换;环棱螺的存在会影响水体中氮形态及藻类的转化^[3];沉水植物能有效吸收和利用水体中的营养盐^[4]. 另一方面,环棱螺与沉水植物具有明显的互利关系^[5],螺-草(沉水植物)系统对于退化水环境及生态系统的修复具有重要的生态调控功能^[6],且具有良好的水质净化效能^[7].

根据笔者对固城湖围垦区池塘河蟹养殖投饲利用情况调查估算,每日投放的未被摄食利用饵料相当于每平方米水面日投入25 mg 氮和5 mg 磷,按池塘平均水深1 m 计,水体氮磷浓度每日将分别增加0.025 mg·L⁻¹和0.005 mg·L⁻¹.由此累积,池塘水体氮磷浓度将会达到很高的水平.同时池塘中螺类

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代谢也将排出大量的氨氮及磷酸盐^[8~11].而水质的跟踪监测表明,池塘水体的氮磷浓度平均约为 1.5 mg·L⁻¹和 0.05 mg·L⁻¹,可见未被摄食饵料的氮磷及水生动物代谢产物的氮磷主要归宿不是水体,而

及水生动物代谢产物的氮磷主要归值不是水体,而是沉积至底泥或被沉水植物、浮游植物、微生物等转化利用. 笔者的研究表明投螺种草的河蟹养殖池塘中水草(沉水植物)收割是氮磷营养盐输出的主要途径,然而池塘螺-草净化系统中水体氮磷营养盐的归趋是否与此结论一致还不得而知.

稳定同位素示踪是一种用同位素或其标记化合 物指示和追踪相应元素或化合物在生物体及其环境 介质中迁移、转化和积累的方法. ¹⁵N同位素添加技 术已经广泛应用到水生态系统中用于示踪氮素的吸 收、迁移、转化规律和消费者食物来源的分析 中[12,13]. 其中,定量添加15N同位素,通过测定目标 组分中¹⁵N原子丰度,从而得出水生态系统中各生物 ¹⁵N的利用量和转移效率越来越受到重视. Dodds 等[14]将15NH4Cl添加到河流生态系统中,研究了外 源氮在食物网各营养级生物的分配量和转移效率. 通过原位标记实验,Barrón等[15]发现地中海有海草 (Posidonia oceanica)的沉积物比无海草的沉积物更 能有效地滞留藻类来源的氮. Zhang 等[16]将15N标记 的微囊藻注射到沉积物,研究不同大型植物吸收 N 的能力,结果表明苦草系统滞留了47.9%的氮,大 于伊乐藻滞留的31.6%,从而揭示了沉积在太湖岸 边沉积物中的蓝藻微囊藻碎屑是大型沉水植物生长 所需要的一个重要的营养来源.

因此本实验构建模拟池塘的螺-草净化系统,利

用生物分析手段和稳定同位素示踪技术研究河蟹养殖池塘螺-草系统中环棱螺和苦草自身的生长及其作用下氮素环境的归趋,以揭示螺-草系统改善池塘水质的机制,并为该系统优化提供理论参考.

1 材料与方法

科

1.1 实验材料

实验基地高淳县阳江镇狮树水产项目渔场为固城湖围垦区.实验容器为直径 25 cm、容积 15 L的塑料桶,塑料桶均匀放置在露天场地上,保证受光均匀.选用养殖池塘最常见的苦草(Vallisneria spiraslis)和环棱螺(Bellamya)分别作为实验的沉水植物和底栖动物. 苦草颜色鲜绿,叶片数 4~6 片,植株长 20~25 cm,环棱螺个体大小均匀,清洗表面附着物,吸干、称重,相关指标见表 1 和表 2.实验自 2010-07-07~2010-08-14,持续 38 d.

实验用水为经 500 目筛绢网过筛后的池塘水,每个实验桶注入 14 L,然后添加浓度为 10 mg·L⁻¹的¹⁵NH₄Cl(99% ¹⁵N)溶液 5 mL,搅拌均匀,添加的¹⁵NH₄Cl量很小,不会对水体的氨氮浓度造成影响.沉积物底泥为池塘底泥,露天晒干后,研磨过 60 目的不锈钢筛网.沉积物沙子为江滩中的细沙,自来水冲洗干净,露天晒干后,过 60 目不锈钢筛网.

表 1 螺的初始质量(湿重)和个数

Table 1 Initial	mass(wet	weight)	and amou	nt of ex	perimenta	l snails	
项目	A1	A2	A3	D1	D2	D3	
质量(湿重)/g	25. 86	26. 5	26. 25	26	26. 37	26. 04	-
数量/个	14	14	14	14	14	14	

表 2 苦草初始质量(湿重)

Table 2 Initial mass(wet weight) of V. spiralis

项目	A1	A2	A3	B1	B2	В3	D1	D2	D3	E1	E2	Е3
质量(湿重)/g	13.41	13. 37	13. 33	13. 36	13. 49	13. 34	13. 27	13.5	13. 6	13.4	13. 35	13. 48

1.2 实验设计

实验设置以底泥和沙子为基质的两组实验,每组实验设置2个处理组1个对照组,每组设置3个重复.两个处理组为苦草和苦草+螺,对照组为无草无螺.为方便描述,将底泥组的苦草+螺处理组简称为A,苦草处理组为B,对照组为C,沙子组分别为D、E和F.

按实验设计(表 1 和表 2)放入苦草和螺,每个实验桶里分别加入预处理过的质量(干重)为 1.5 kg 的池塘底泥和沙子,缓慢注水,尽量不对底泥造成扰动,减少底泥悬浮及营养盐的释放.

1.3 样品采集及分析方法

桶内水体静置 3 h 后采集原水样,以后定期采集水样,每次在实验桶水下 10 cm 处用导管虹吸采集样品 200 mL,其中将 50 mL 水样经过 GF/C 醋酸纤维膜过滤.同时采集初始苦草、环棱螺、底泥样品,用塑封袋包装好待预处理及分析.实验结束后采集底泥组的各个处理及对照组的表层底泥(约 2 cm),用塑封袋包装.同时测量各处理组的苦草及环棱螺物理指标,并采集样品测定其氮磷含量及稳定同位素分析.

水质指标测定:未经过滤水样测其总氮(TN)和

总磷(TP),经 GF/C 滤膜过滤的水样,测其氨氮 (NH_4^+ -N). TN、TP 采用过硫酸钾消解法同时测定, NH_4^+ -N采用纳氏试剂法测定. 测定方法参照文献 [17].

样品氮磷含量测定:采集的苦草(洗净表层的附着物及根部的底泥或沙子)、螺(取螺肉部分)及底泥样品放烘箱中于40℃下烘48 h,烘干后测定苦草的地上部分及地下部分的质量(干重),然后将全部样品用玛瑙碾磨磨碎,经100 目筛过筛,用样品袋装好后待测. TN含量分析方法为过氧化氢-硫酸消化-凯氏定氮法,TP含量分析方法为过氧化氢-硫酸酸溶-钼锑抗比色法. 测定方法参照文献[18].

样品氮稳定同位素测定:采集的苦草及螺肉样品清洗干净,洗去表层粘附的 $^{15}NH_4^+$ -N,然后放冷冻干燥机干燥 48 h,干燥后放玛瑙碾磨中磨碎装入塑料管中. 然后称取一定质量的样品放入同位素质谱仪测定氮稳定同位素值. 样品的氮稳定同位素成分用 Thermoelectron Flash 1112 元素分析仪连接一个Thermoelectron Delta V 质谱仪分析(EA-IRMS). $\delta^{15}N$ 的测定误差小于 0.3%.

1.4 过量¹⁵N 计算

¹⁵N 数据是指每 g 干物质¹⁵N的 μmol,根据文献 [16]中所阐述的公式进行计算:

过量
$$^{15}N(\ \mu\mathrm{mol/g}) = \left\{ \left[\frac{(\ at\%^{\ 15}N_{_{\xi H_{-}}} \ at\%^{\ 15}N_{_{\otimes \mathbb{M}}})}{100} \right] \times \right.$$

[样品中氮含量] }/[样品的质量(干重)] (1)

at% ¹⁵
$$N_{\text{#H}} = \frac{100 \times R_{\text{+}\%} \times [(\delta^{15}N_{\text{#H}}/1\ 000) + 1]}{[1 + R_{\text{+}\%} + R_{\text{+}\%}(\delta^{15}N_{\text{#H}}/1\ 000)}$$

(2)

$$\delta^{15}N(\%o) = [(R_{\#_{loc}}/R_{+\%}) - 1] \times 1000$$
 (3)

其中,at% $^{15}N_{\text{sm}}$ 代表第 0 天原子丰度值; $R_{\text{#}\text{H}}$ = $^{15}N/^{14}N$ (样品的比率); δ ^{15}N 相对于大气氮比率的过量的值, R_{t} = 0.003 676 5.

1.5 数据分析方法

利用 SPSS 16.0 进行数据统计分析,用一元方差分析不同处理组的实验数据差异性,并用 Duncan 方法进行组间差异分析. 利用 Sigmaplot 10.0 进行作图分析.

2 结果与分析

2.1 水体营养盐变化

由图1可知,底泥组和沙子组的各个处理总氮、 总磷和氨氮浓度随时间的变化趋势均基本一致,底 泥组呈现先降后增再降的波动,而沙子组基本为逐渐下降,最终均稳定在较低水平. 2 组均在前 5 d 下降最快,底泥组的总氮、总磷的下降幅度为 53%和 63%,均分别大于沙子组的 49%和 55%,而氨氮的下降幅度为 61%,小于沙子组的 71%.实验结束后,底泥组各个处理的总氮、总磷和氨氮均值为1.10 mg·L^{-1} 、0.052 mg·L^{-1} 、0.30 mg·L^{-1} 、0.18 mg·L^{-1} ,而 2 组的 3 个处理间总氮、总磷的差异均不显著(P > 0.05),沙子组的 D、E 处理氨氮浓度却显著高于对照组(P < 0.05).

2.2 苦草生长指标的比较

由图 2 可知,实验结束后苦草的质量(湿重)较 初始值显著增加,底泥组的苦草处理及螺-草处理的 质量(湿重)均显著高于沙子组,A、B、D、E 这4个 处理间均差异显著(P<0.05),A处理最高,增加了 580%, E 处理最低, 增加了66%. 苦草的株数也显 著增加,其中 A 处理平均增加了 6.6 株,显著高于 B 处理的 3.6 株, 而 D 处理和 E 处理间差异则不显著 (P>0.05). 由图 3 可知, A、D 处理的地上部分与 地下部分质量(干重)均高于 B、E 处理,其中 A 和 B 处理间差异显著(P < 0.05), 而 D 和 E 处理间差 异不显著(P > 0.05),且底泥组的2个处理地上部 分质量(干重)均显著高于沙子组相应处理(P < 0.05),而地下部分质量(干重)则不受基质类型的 影响. A 处理的苦草地上部分与地下部分质量(干 重) 比显著低于 B 处理(P<0.05), 而 D 和 E 处理 间差异不显著(P>0.05). 基质类型对苦草的含水 率有较显著的影响, A 和 B 处理的苦草含水率均分 别显著高于 D 和 E 处理(P < 0.05).

2.3 各要素氮磷含量的比较

由图 4 可知,实验结束后 4 个处理的苦草地上部分及地下部分氮磷含量均较初始有所下降,且地上与地下部分下降幅度基本一致,其中沙子组的 D、E 处理的苦草地上部分总氮和地下部分总磷含量降幅最大,平均分别下降了62%、67%、72%和66%,且 D、E 处理的苦草地下部分总磷含量降幅均显著大于 A、B 处理(P < 0.05).

由图 5 可知, 螺肉的总氮总磷含量变化较小, A、D 处理间无显著差异, 与初始值差异也不显著 (*P* > 0.05). A、B、C(对照)处理间底泥的有机质、总氮、总磷含量差异均不显著(*P* > 0.05).

2.4 ¹⁵N丰度及收支平衡分析

由图6可知,实验结束后苦草、环棱螺和底泥

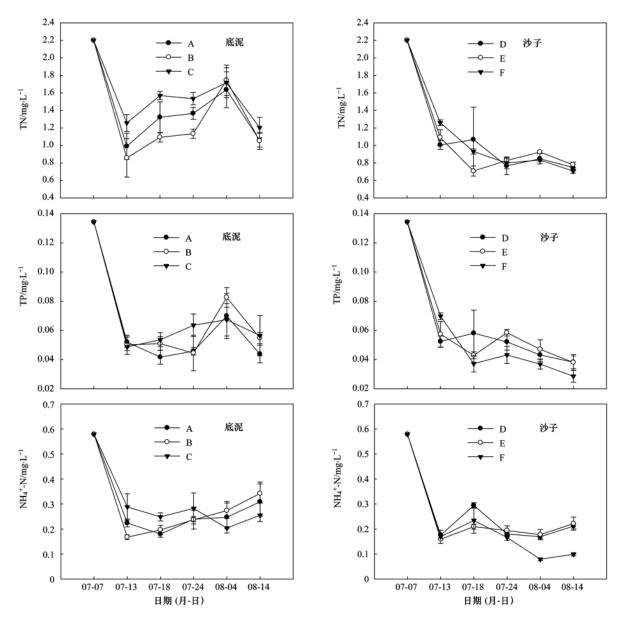


图 1 底泥组和沙子组水体营养盐的变化

Fig. 1 Changes of nutrient in water of sediment groups and sand groups

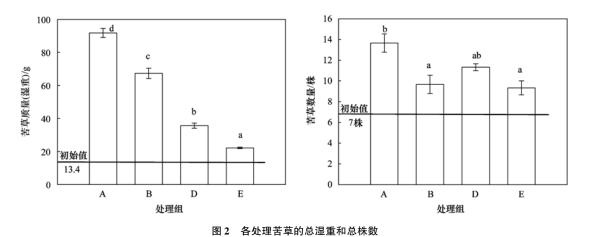


Fig. 2 Total wet weight and ramet amount of V. spiralis in different treatments

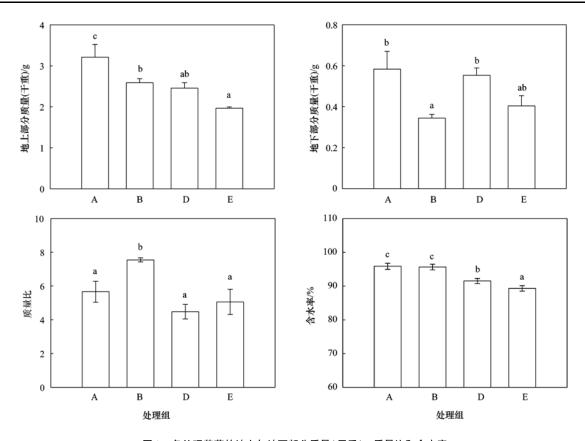


图 3 各处理苦草的地上与地下部分质量(干重)、质量比和含水率

Fig. 3 Dry weight, mass ratio and moisture content of aboveground and underground parts of V. spiralis in different treatments

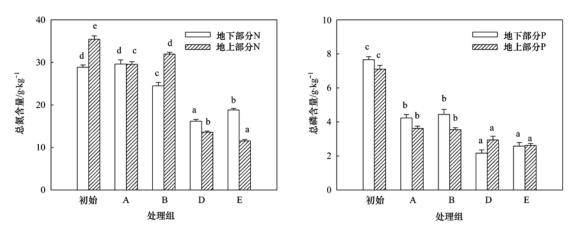


图 4 各处理苦草地上部分和地下部分的氮磷含量

Fig. 4 Nitrogen and phosphorus content of aboveground and underground parts of V. spiralis in different treatments

均对氮有所吸收和累积, 15 N丰度显著高于初始值,其中 A、D 处理的苦草地上部分 15 N丰度分别显著低于 B、C 处理(P < 0.05),C 处理(对照)的底泥 15 N丰度低于 B 处理、高于 A 处理,且处理间差异显著(P < 0.05).由图 7 可知,在实验结束时 A 处理的苦草根部吸收储存了 1.07%的 15 N,苦草茎叶吸收储存了 7.74%的 15 N,环棱螺吸收较少,只占 0.06%,底泥滞留了 5.73%的 15 N; B 处理的苦草根部吸收储存了 0.45%的 15 N,苦草茎叶吸收储存了 17.44%的 15 N,底

泥滯留了 38.77% 的 15 N. D 处理苦草根部吸收储存了 1.84% 的 15 N, 苦草茎叶吸收储存了 0.11% 的 15 N, 环棱螺吸收也较少, 只占 0.14%; E 处理的苦草根部吸收储存了 0.94% 的 15 N, 苦草茎叶吸收储存了 0.17% 的 15 N.

3 讨论

水体中悬浮颗粒物的自然沉降及沉积物短时间内的吸附现象较为明显^[19,20].实验前 5 d 内底泥组

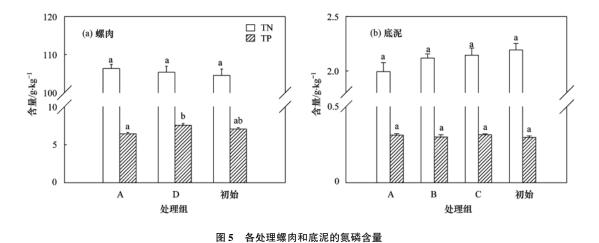


Fig. 5 Nitrogen and phosphorus content of snail muscle and sediment

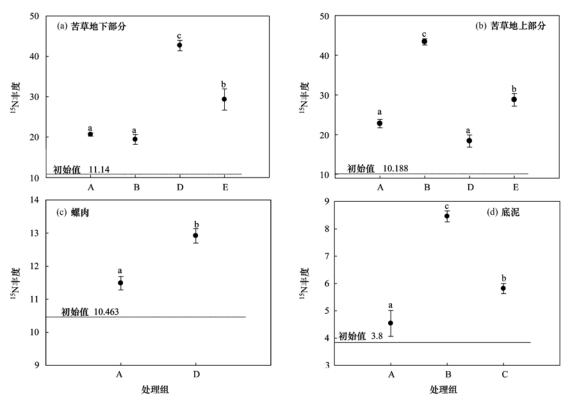


图 6 苦草、环棱螺和底泥¹⁵N丰度

Fig. 6 15 N richness of V. spiralis, Bellamya and sediment

和沙子组的各处理水体颗粒态氮磷自然沉降及沉积物对溶解态氮磷的吸附,使水体的总氮、总磷和氨氮浓度均有大幅度下降,而实验结束后底泥组各处理的氮磷含量均显著高于沙子组,可能与底泥中氮磷的少量释放有关. 沉水植物对水体中氮磷的净化作用主要表现在同化吸收^[21]、改善根系微生态环境从而增进微生物的生化作用^[22,23]并降低沉积物中氨氮和硝态氮的扩散通量^[24]、促进沉积物对上覆水中磷的吸附^[25]、抑制沉积物-水界面氮磷的释放^[24,26]等. 环棱螺主要通过呼吸、摄食作用促进悬

浮颗粒物沉降^[27],同时其爬动行为对沉积物-水界面有较大的影响,提高了沉积物中底泥硝化及反硝化速率,加快沉积物中的氮循环^[28,29].而本研究的苦草和环棱螺的净化作用不明显,实验结束后2个基质组的3个处理间总氮、总磷的差异均不显著(P>0.05),主要原因为实验水体溶解态氮磷浓度较低,随着颗粒态氮磷沉降后,水体氮磷已达到较低水平,环棱螺与苦草存在净化阈值,且高温时少量苦草浮叶腐败分解以及环棱螺自身代谢释放氮磷,因此水体氮磷稳定在较低的水平,处理组与对照组差异

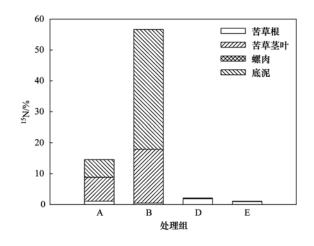


图 7 各个处理各要素过量¹⁵N比例

Fig. 7 Proportion of excess ¹⁵N of each element in different treatments

不显著.

螺类对水生植物茎叶的牧食减少了植物的光照 限制及其附着生物的营养盐竞争等影响,促进了水 生植物的生长[9],水生植物也为螺类提供了牧食产 卵的基质条件及躲避捕食者的栖息场所[30]. 不同 密度的环棱螺均显著促进受附植物苦草的生长,螺-草的互利关系成立[5]. 本实验结果同样也表明环棱 螺的存在显著促进了苦草分株和根系、茎叶的生 长,而实验环棱螺密度较低,通过实验中观察可知环 棱螺大多爬行于底泥表层或钻入底泥,对苦草叶片 表层的附着生物的刮食较少,因此本研究环棱螺促 进苦草生长的主要机制不是通过牧食苦草茎叶的附 着物,而是环棱螺爬行、钻孔等运动增大了沉积物 含水率、孔隙度,提高了沉积物中总微生物活 性[31],改善了根系生境,促进根系向深层扩展,从而 吸收更多的营养物质,然后转移到茎叶,促进苦草的 分株、新芽的萌发及茎叶的生长,且在营养盐丰富 的基质中此种促进效应更显著,同时环棱螺自身代 谢释放也为苦草生长提供了营养[11]. 沉积物是很 大的氮磷库,根系系统是沉水植物吸收氮磷营养盐 的主要部位[32],因此有机质含量丰富的底泥组苦草 的生长速率和含水率均显著高于沙子组,陈开宁 等[33]的研究也有相似的结论,但基质类型不会显著 影响苦草的分株. A、B 处理的苦草地上和地下部 分的总氮含量与初始值差异较小,但分别显著高于 D、E处理,而A、B、D和E处理的苦草地上和地下 部分的总磷含量都显著低于初始值,可能由于根系 主要从沉积物中吸收氮,对磷的吸收相对较少,根系 中的氮磷输送到茎叶中,以提供茎叶萌发分株所需 要的营养,同时氮通过叶片的渗透压力向水体释放以保证植物系统的物质浓度平衡^[21],特别是水体营养盐较低时,会继续分泌氮磷以与水体达到平衡^[34].

沙子组苦草只能从水体中吸收利用溶解态氮, 稳定同位素示踪结果显示,D、E处理的苦草地上与 地下部分吸收储存少量的¹⁵N,最大仅为1.84%,这 也证实了苦草对水体中溶解态氮的吸收较少,最大 只有 10% [22]. D 处理的苦草地下部分 15N 丰度显著 高于 E 处理, 表明环棱螺能促进苦草根系吸收水体 中的氨氮. B 处理底泥的¹⁵N丰度显著高于 C 处理, 且 B 处理的底泥滞留了 38.77% 的¹⁵N, 苦草茎叶吸 收储存了 17.44% 的¹⁵N,表明静态水体中的大部分 溶解态氮通过表层渗透及底泥吸附进入底泥.且同 时苦草有较高的从底泥富集营养盐的能力[32,35].一 方面根系从底泥中吸收生物可利用的氨氮,并随生 物量增加间隙水中游离的氨氮含量减少,另一方面, 苦草生长光合作用可增加上覆水体溶解氧.氧可部 分向沉积物-水界面下扩散,并通过茎、叶和根系向 沉积物输送,使沉积物氧化还原电位升高[25],促进 氮的硝化、抑制下层沉积物氨化,使间隙水中的氨 氮浓度逐渐降低[36],因此泥-水界面存在浓度梯度, 使上覆水中氨氮逐渐向底泥中渗透,从而提高了底 泥滞留氨氮的能力. 环棱螺通过改变沉积物表层的 结构、增加沉积物表层的含氧量和含水率以及增强 总微生物活性[31,37],促进沉积物氨氮的释放(包含 自身代谢释放)和对上覆水 NO; 的吸收,提高了硝 化和反硝化速率,并增加了沉积物铁结合态磷含量, 从而促进系统中氮磷的循环. 因此环棱螺的存在反 而使 A 处理的底泥仅滞留了 5.73% 的¹⁵N,苦草茎叶 吸收储存了 7.74% 的¹⁵N,均远小于 B 处理. 综上分 析可得. 螺-草系统中氮素的环境归趋模式为: 水体 外源性氨氮及环棱螺自身代谢释放的氨氮少量由沉 水植物直接同化吸收,主要通过泥-水界面进行迁移 转化,大部分被沉水植物根系吸收,少部分通过硝 化/反硝化作用去除,其余则沉积在底泥;系统中的 大部分氮素以沉水植物的途径输出,环棱螺的存在 通过增强泥-水界面硝化反硝化过程及促进沉水植 物的生长而加快水体及底泥中氮素的输出. 笔者对 河蟹养殖池塘的污染负荷输出研究表明,底泥中氮、 磷的平均累积量分别占总输入的29%和41%,沉水 植物从池塘携带的氮、磷总量分别占总输出的86% 和88%,这也佐证了以上的归趋模式. 本实验进行 预处理已排除其它底栖动物的干扰,且实验中观察

发现浮游藻类和浮游动物生物量较小,因此水体中其它生物对实验结果的影响较小.由于实验条件的限制未测定水体¹⁵N的值,无法计算硝化-反硝化过程去除的氮,因此有待进一步深入研究,以了解系统中氮素的迁移转化过程.同时池塘中未被摄食的饵料是池塘系统中很大的营养源,研究其对水体营养盐的贡献及其在系统中迁移转化对池塘水质调控具有重要的意义.

池塘生态养殖系统中通过苦草和环棱螺的相互作用减少了外源性氮磷在水体和底泥中的累积,从而使养殖水体营养盐保持在较低的水平. 然而实际养殖中也发现,螺-草系统的构建需要根据不同时期控制一定的比例,养殖初期如投放过多环棱螺牧食会对幼嫩植物造成危害而不利生长,养殖中后期过高则会因其代谢释放造成水体营养盐升高,而促进附着生物^[38,39]和浮游藻类^[3]的生长,造成水体溶解氧下降,导致环棱螺及沉水植物生长限制甚至死亡^[40],使水体恶化. 因此在养殖的不同阶段合理配置螺-草结构是整个养殖过程中水质调控的关键.

4 结论

- (1)环棱螺主要通过改善泥水界面促进了苦草根系扩展、叶片分株及茎叶生长,其中以底泥为基质表现更显著,实验结束后苦草总湿重和分株数较初始增加了580%和6.6株.
- (2) 苦草对水体中溶解态氮的同化吸收较少, 底泥是苦草生长的主要营养源,水体中氨氮通过泥水界面进入底泥,其大部分被沉水植物根系吸收,少部分通过硝化/反硝化作用去除,其余则沉积于底泥,同时苦草能增强底泥对氨氮的滞留能力.
- (3)螺-草净化系统中,苦草是氮素去除的最终 载体,环棱螺的存在通过促进苦草生长及加强泥-水 界面硝化和反硝化作用来加快系统中氮素的去除.
- (4)利用稳定同位素示踪技术研究池塘氮素环境归趋模式,能深入揭示螺-草系统的水质净化机制,并为养殖池塘螺-草水质净化系统优化及养殖过程污染控制提供理论依据.

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