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土壤质地对自养固碳微生物及其同化碳的影响

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摘要:自养微生物可同化大气中的 CO₂ 并将其转化为土壤有机碳,对提高农田土壤的碳吸收和碳储存有重要意义,然而土壤质地对自养固碳微生物功能种群及其同化碳的影响机制还不清楚. 本研究选取亚热带地区同一母质发育而成的两种质地水稻土壤(壤质黏土和砂质黏壤土),通过¹⁴C-CO₂连续标记技术结合室内模拟培养实验,探讨土壤质地对自养微生物同化碳(¹⁴C-SOC)、自养微生物截留碳(¹⁴C-MBC)和自养微生物可溶性碳(¹⁴C-DOC)的影响. 以固碳功能基因(cbbL 基因)作为指示基因,结合 PCR 和克隆测序技术,分析不同质地土壤自养固碳微生物群落结构和多样性的差异. 结果表明,壤质黏土¹⁴C-SOC、¹⁴C-MBC和¹⁴C-DOC平均含量分别为 133.81、40.16 和 8.10 mg·kg⁻¹,均显著高于砂质黏壤土 ¹⁴C-SOC(104.95 mg·kg⁻¹)、¹⁴C-MBC(33.26 mg·kg⁻¹)和 ¹⁴C-DOC(4.18 mg·kg⁻¹)平均含量(P<0.05),说明土壤质地显著影响了土壤自养微生物碳同化量以及自养微生物同化碳在土壤中的转化. 稀疏曲线、细菌 cbbL 基因文库覆盖度以及多样性指数分析结果显示壤质黏土固碳细菌群落多样性高于砂质黏壤土. 系统发育分析表明,壤质黏土细菌 cbbL 基因序列与 Rhodoblastus acidophilus、Blastochloris viridis、Thauera humireducens、Mehylibium sp.、Variovorax sp. 等具有一定的同源性,而砂质黏壤土 cbbL 基因序列主要与根瘤菌和放线菌同源. 可见,土壤质地对自养固碳微生物群落结构和多样性产生了深刻的影响,壤质黏土中较高的黏粒含量、土壤养分含量和阳离子交换量可能有利于维持更高的自养固碳微生物多样性和活性,从而导致不同质地土壤自养微生物碳同化量及其转化存在显著差异.

关键词:土壤质地;固碳自养微生物; CO_0 ,同化;cbbL基因;多样性;群落结构

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Effects of Soil Texture on Autotrophic CO₂ Fixation Bacterial Communities and Their CO₂ Assimilation Contents

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Abstract: Autotrophic bacteria can assimilate atmospheric carbon dioxide (CO₂) and convert CO₂ into organic carbon. The CO₂ fixation by autotrophic bacteria is important for the improvement of carbon sequestration in agricultural soils. However, the effect of soil texture on autotrophic CO2 fixation bacteria and their CO2 fixation capacity is still unknown. Here, two paddy soils with different textures (loamy clay soil and sand clay loam soil) were incubated with continuous 14C-CO, in a glass chamber. The two soils were developed from the same parent. At the end of 110 days incubation, the ¹⁴C-CO₂ incorporated in soil organic carbon (¹⁴C-SOC), microbial biomass carbon (14C-MBC) and dissolved organic carbon (14C-DOC) were measured to explore the effects of soil texture on the autotrophic bacterial CO2 fixation rates. The effect of soil texture on the composition and diversity of autotrophic CO2 fixation bacterial community was investigated using cloning and sequencing of the cbbL gene, which encodes ribulose-1,5-biphosphate carboxylase/oxygenase (RubisCO) in the Calvin cycle. The results showed that the average contents of 14C-SOC, 14C-MBC and ¹⁴C-DOC were 133.81, 40.16 and 8.10 mg·kg⁻¹ in loamy clay soil, respectively, which were significantly higher than their corresponding contents in sand clay loam soil (P < 0.05). This suggested that soil texture not only affected the amounts of autotrophic bacteria CO₂ fixation but also had an effect on the transformation of microbial assimilated ¹⁴C in soil. The cbbL gene libraries of two soils were significantly different as revealed by libshuff analyses (P < 0.05). Phylogenetic analysis showed that cbbL sequences from the loamy clay soil were closely affiliated with known cultures such as Rhodoblastus acidophilus, Blastochloris viridis, Thauera humireducens, Mehylibium sp. and Variovorax sp., whereas these sequences belonging to the sand clay loam soil were related to branching lineages originating from Rhizobiales and Actinomycetales. Rarefaction curve, clone library coverage and diversity index

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analysis based on bacterial *cbbL* clone libraries indicated that the loamy clay soil had higher *cbbL* gene diversity compared to the sand clay loam soil. These results suggested that soil texture had a pronounced effect on the composition and diversity of autotrophic CO₂ fixation bacterial communities. The higher clay content, nutrient availability and cation exchange capacity may stimulate the growth and activity of autotrophic bacteria, and result in the higher amounts of ¹⁴C in loamy clay soil. These data broaden the understanding and knowledge of mechanisms of microbial carbon fixation and their influencing factors in agricultural soils.

Key words: soil texture; autotrophic bacteria; CO2 assimilation, cbbL gene; diversity; community structure

自养微生物在农田土壤中广泛分布,它们具有 同化 CO, 并将其转化为土壤有机碳的能力,对调节 大气中 CO,浓度和提高农田土壤的碳固定有着重要 意义[1,2]. 卡尔文循环(Calvi-Bussham cycle)是自养 微生物同化 CO, 的主要途径,其中核酮糖-1,5-二磷 酸梭化酶/加氧酶(RubisCO) 是卡尔文循环中的关 键酶,由 cbbL 基因编码[3]. 近年来,利用 cbbL 基因 对农田土壤自养微生物及其碳同化过程的研究已取 得较大进展. 结合碳同位素示踪实验,研究者发现 参与农田土壤碳同化的主要是兼性自养微生物,且 农田土壤自养微生物多样性、群落结构及其碳同化 量受土壤类型[2,4,5]、施肥措施[6,7]、土壤深度[8]、 根际效应[9]和耕作制度[10]等多种因素的影响. 这 些因素通过改变土壤的物理化学性质而影响土壤自 养微生物多样性和群落结构,进而对土壤自养固碳 微生物介导的碳同化过程产生影响.

土壤质地是土壤较为重要的物理特性之一,一方面,土壤质地可直接影响土壤的孔隙状况,进而影响土壤的透光率和通气性^[11];另一方面,土壤质地还会影响土壤的养分状况和水分含量,从而影响微生物的生存环境及其代谢活性^[12,13].土壤通气透水以及土壤肥力水平是影响自养微生物碳同化过程的重要生态因子^[1,6,14],不同质地土壤中这些生态因子的差异可能对自养微生物群落结构及其碳同化过程产生重要影响,然而,目前有关土壤质地对自养微生物多样性、群落结构及其碳同化量影响的研究还鲜

见报道. 因此,本研究选取亚热带地区同一母质发育而成的两种质地水稻土壤(壤质黏土和砂质黏壤土),采用¹℃-CO₂连续标记技术结合室内模拟实验,量化不同质地土壤自养微生物同化碳(¹℃-SOC)、自养微生物截留碳(¹℃-MBC)和自养微生物可溶性碳(¹℃-DOC)的差异;同时采用分子生物学技术,基于细菌 cbbL 基因对土壤自养微生物群落结构和多样性进行分析,通过揭示不同质地稻田土壤自养微生物碳同化的机制,以期为更深入全面了解稻田土壤微生物固碳机制及其影响机制提供理论和数据支持.

1 材料与方法

1.1 供试土壤与前处理

供试土壤于 2010 年 10 月采自湖南省长沙县干杉镇水稻田,共采集壤质黏土和砂质黏壤土 2 种质地土壤,均为河流冲积物发育的水稻土. 采用直径为 5 cm 的不锈钢土钻采集 0~20 cm 耕作层土壤,去除石块和根系后运回实验室风干. 风干土壤过 0.25 mm筛,混匀后分为两部分,一部分用于测定土壤基本理化性质(用于 SOC 测定的土壤过 0.149 mm 筛);剩余土壤用蒸馏水调节含水量至饱和田间持水量,分装于直径为 10 cm、高为 20 cm 的 PVC 盆钵,每钵装入量为 1.00 kg(以烘干后的质量计),装入高度为 17 cm,每种土壤设置 4 个重复,在 25℃ 预培养 14 d. 两种供试土壤的基本理化性质见表 1.

表 1 供试土壤基本理化性质

Table 1 Physical and chemical characteristics of the investigated soils

•	机械组成			理化性质				
土壤质地	砂粒含量	砂粒含量 粉砂粒含量 黏粒含量 pH		SOC	TN	TP	CEC	
	/%	/%	/%	pii	∕g•kg ⁻¹	/g•kg ⁻¹	/g•kg ⁻¹	∕emol•kg ⁻¹
壤质黏土	39. 04	21. 20	39. 76	6. 15	21. 89	2. 64	1. 05	11. 79
砂质黏壤土	59. 65	22. 06	18. 29	5. 09	17. 03	2. 52	0. 58	5. 6

1.2 ¹⁴C-CO,连续标记培养实验

将预培养后的装盆土壤转移至 14 C-CO₂标记箱进行连续标记培养,培养装置和方法参考 Ge 等 $^{[15]}$ 建立的方法,连续标记培养时间为 110 d. 14 C-CO₂由 14 C-NaHCO₃(1 mol·L $^{-1}$,16.5×10 3 Bq·mL $^{-1}$) 和 HCl

(1 mol·L⁻¹)反应生成,通过调节¹⁴C-NaHCO₃ 和 HCl 溶液的加入量和加入次数使标记箱内 CO₂ 浓度维持在 350 μ mol·mol⁻¹. 标记箱内温度白天为 31℃ ±1℃,夜间为 24℃ ±1℃,光照时间为 08:00 ~ 20:00(时间形式修改正确),相对湿度为 80% ~

90%,光照强度约为 500 $mmol \cdot (m^2 \cdot s)^{-1}$ PAR. 标记培养过程中,及时补充去离子水维持土壤水层深度 1~2 cm.

110 d 连续标记培养结束后,采集 PVC 盆钵内供试土壤样品并充分混匀. 一部分立即处理,用于含水率、¹⁴C-MBC和¹⁴C-DOC的测定;一部分室内自然风干后过 0. 149 mm 筛,用于¹⁴C-SOC含量的测定;一部分(约 100 g)用液氮速冻后转移至 – 80℃ 冰箱,用于固碳自养微生物群落组成分析.

1.3 测定和分析方法

1.3.1 土壤基本理化性质测定^[16]

以水为浸提剂,采用水土比 2.5:1测定土壤pH;土壤阳离子交换量采用 EDTA-铵盐快速滴定法测定;土壤机械组成采用吸管法测定;土壤有机碳(SOC)和全氮(TN)采用碳氮元素分析仪(VARIO MAX,Germany)测定;土壤全磷采用碱熔-钼锑抗分光光度计法(UV-2450,Japan)测定.

1.3.2 土壤⁰ 放射性强度测定

土壤总有机碳中¹⁴C放射性强度采用 Wu 等^[17] 方法: 称取 1.5 g 土壤样品至双颈烧瓶中,向烧瓶加入 20 mL 重铬酸钾 (0.2 mol·L^{-1}) 溶液和体积比为 5:1的浓硫酸-浓磷酸混合液(30 mL)作为氧化剂, 165 © 回流消化 8 min,随后持续向双颈烧瓶中通入 0_2 ,继续消化 10 min,消化过程中产生的 $C0_2$ 经分离纯化后用 40 mL NaOH 溶液充分吸收 (0.4 mol·L^{-1}) . 吸收液中¹⁴C放射性强度 $(^{14}\text{C-SOC})$ 采用液体闪烁仪(LS-6500,Beckman)测定. $^{14}\text{C-SOC}$ 含量计算方法参见文献[18].

¹⁴C-MBC和¹⁴C-DOC含量测定参考 Wu 等^[19]建立的氯仿熏蒸提取-碳自动分析仪法进行: 称取 4 份 10.00 g 新鲜土壤,其中两份按 1:4的土水比加入 K₂SO₄ 浸提液(0.5 mol·L⁻¹),另两份在真空干燥器 内用氯仿熏蒸 24 h,去除氯仿后立即浸提. 浸提液中 ¹⁴C 放射性强度采用液体闪烁仪(LS-6500, Beckman)测定,根据不熏蒸土样浸提液中 ¹⁴C 放射性强度计算 ¹⁴C-DOC含量,根据熏蒸土样与不熏蒸土样浸提液 ¹⁴C 放射性强度之差计算 ¹⁴C-MBC含量,计算方法参见文献[2,18].

1.3.3 土壤细菌 cbbL 基因克隆、测序和系统发育分析

采用 FastDNA 土壤试剂盒(Mpbio, USA)分别提取壤土和砂土 DNA,并利用紫外分光光度计(Nanodrop, Germany)测定 DNA 的浓度和纯度. 采用引物 K_{2f}(5'-ACCA[C/T]CAAGCC[G/C]AAGCT

「C/G]GG-3')和 V_{2r}(5'-GCCTTC[C/G]AGCTTGCC 「C/G]ACC[G/A]C-3')分别扩增细菌 cbbL 基因片 段^[20]. PCR 扩增反应体系配置如下: 12.5 μL 2× PCR MasterMix(天根,中国), DNA 模板约 50 ng,上 下游引物各 $0.1 \, \mu \text{mol} \cdot \text{L}^{-1}$, 无菌水补至 $25 \, \mu \text{L}$. 扩增 程序采用梯度降落 PCR 程序,反应条件如下: 95℃ 预变性 3 min,5 个循环为 95℃ 变性 30 s,66~62℃ 退火 50 s,72℃ 延伸 90 s,每次循环退火温度降低 1℃;后30个循环为95℃变性45 s,62℃退火50 s, 72℃延伸90 s; 72℃ 最终延伸10 min. PCR 扩增 产物用1.2%琼脂糖电泳检测,采用琼脂糖凝胶 DNA 回收试剂盒(天根,中国)回收目标片段,方 法按说明书进行. 将 PCR 回收产物与 pGEM-Teasy (Promega, 德国) 载体连接后, 转入 E. coil DH5α 感 受态细胞,构建重组质粒,通过蓝白斑筛选,随机 挑选白色菌落,采用载体通用引物 SP6 和 T7 鉴定 阳性克隆,筛选含目标片段的克隆送至华大基因 公司测序,分别建立壤土和砂土细菌 cbbL 基因文 库. 利用 Mothur 软件对所获得的 cbbL 基因序列进 行分析,将相似性大于95%的序列归为同一个操 作单元(OTUs). 将 OTU 的代表性核苷酸序列翻 译为氨基酸序列,在 NCBI(http://www.ncbi.nlm. nih. gov/blastx)中进行同源性比对,挑选相似度较 高的同源氨基酸序列,并利用 MEGA 6.0 中的邻 接法(Neighbor-joining)构建系统发育树. 本研究 所得序列在 EMBL 中的登录号为 LT559266-LT559471.

1.4 数据分析

采用 Microsoft Excel 2010 和 SPSS 16.0 对数据进行处理和统计分析,不同质地土壤差异显著性采用独立样本 t 检验. 采用 Mothur 软件分析细菌 cbbL 克隆文库覆盖度(Coverage)、物种丰富度指数(Richness、ACE 和 Chao)和物种多样性指数(Simpson和 Shannon).利用 Mothur软件中的Libshuff命令检验不同质地土壤自养微生物群落组成的差异显著性(P<0.05).

2 结果与分析

2.1 土壤质地对微生物同化碳含量的影响

连续标记培养 110 d 后,不同质地土壤中均检测到了 $^{14}\text{C-SOC}$. 壤质黏土 $^{14}\text{C-SOC}$ 含量范围为 129. 83 ~ 141. 66 mg·kg^{-1} , 砂质黏壤土 $^{14}\text{C-SOC}$ 含量范围为 95. 80 ~ 112. 91 mg·kg^{-1} . 两种质地土壤 $^{14}\text{C-SOC}$ 平均含量差异达显著水平,表现为壤质黏土

高于砂质黏壤土(表 2),说明土壤微生物同化碳含量受土壤质地的影响. 壤质黏土 14 C-MBC和 14 C-DOC含量分别为 37.82~59.35 mg·kg^{-1} 和 7.68~9.02 mg·kg^{-1} ,砂质黏壤土 14 C-MBC和 14 C-DOC含量分别为

29. 47~37. 50 $\text{mg} \cdot \text{kg}^{-1}$ 和 3. 60~4. 79 $\text{mg} \cdot \text{kg}^{-1}$, 壤质黏土¹℃-MBC和¹℃-DOC平均含量均显著高于砂质黏壤土(表 2),表明土壤质地影响了自养微生物同化碳在土壤中的转化过程.

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表 2 不同质地土壤 14 C-SOC、 14 C-MBC和 14 C-DOC含量 $^{1)}$ /mg·kg $^{-1}$

Table 2 Contents of ¹⁴ C-SOC. ¹⁴	⁴ C-MBC and ¹	C-DOC in soils	with different	texture/mg·kg ⁻¹
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	,		0 0
土壤质地	¹⁴ C-SOC 含量	¹⁴ C-MBC 含量	¹⁴ C-DOC 含量
壤质黏土	133. 81 ± 2. 72a	49. 16 ± 4. 81a	8. 10 ± 0. 32a
砂质黏壤土	$104.95 \pm 4.15b$	$33.26 \pm 1.69b$	$4.18 \pm 0.34 b$

1)不同小写字母表示不同质地土壤之间差异显著(P<0.05)

2.2 土壤质地对自养固碳微生物多样性的影响

构建的两种质地土壤样品微生物的 cbbL 基因文库中共获得 335 条有效细菌 cbbL 基因序列. 从表 3 可以看出,以 95% 序列相似度作为 OTUs 划分标准,从壤质黏土和砂质黏壤土中分别获得 106 和 100 个 OTUs. 两种质地土壤细菌 cbbL 基因克隆文库的覆盖度为 50% ~57.8%. 稀疏曲线结果显示各个文库的稀疏曲线趋于平缓,进一步说明本研究建

立的克隆文库具有一定的代表性,能较好地反映土壤中自养微生物多样性(图1). 通过对 Richness、ACE 和 Shannon 等多样性指数进行分析,发现壤质黏土自养微生物多样性高于砂质黏壤土(表3). Libshuff 分析结果显示,壤质黏土和砂质黏壤土细菌cbbL 基因克隆文库存在显著性差异(P < 0.05),说明土壤质地对含 cbbL 基因的自养微生物群落结构造成了显著影响.

表3 不同质地土壤细菌 cbbL 基因多样性

Table 3 Diversity of cbbL-containing bacteria in soils with different texture

土壤质地	克隆子数	覆盖度[8]/%	Richness	ACE	Chao	Simpson 指数	Shannon 指数
壤质黏土	162	50	106	636	278	0. 011 6	4. 425
砂质黏壤土	173	57. 8	100	501	289	0.0117	4. 348

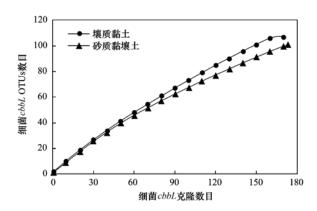


图 1 不同质地土壤细菌 cbbL 克隆文库的稀疏曲线

Fig. 1 Rarefaction curves of bacterial cbbL genes in clone libraries from different texture soils

2.3 土壤质地对自养固碳微生物群落结构的影响

系统发育分析结果显示,本研究获得的 206 个 cbbL 基因型以兼性自养菌(FormIC)为主. 两种质地土壤 cbbL 基因型在第 X cbbL 基因簇分布较均匀,表明不同质地土壤严格自养菌组成差异不大(图 2). 壤质黏土中分别有 23% 和 16% 的 cbbL 基因型分布在第 I 和第 II cbbL 基因簇,与嗜酸柏拉红菌 Rhodoblastus acidophilus、绿色绿芽菌 Blastochloris

viridis、腐殖质还原陶厄氏菌 Thauera humireducens、高效降解菌 Mehylibium sp.、贪噬菌 Variovorax sp.、伯克氏菌 Burkholderiales bacterium 等具有一定的同源性. 壤质黏土中有 28%的 cbbL 基因型为尚未被发现的新 cbbL 基因型,主要分布在第Ⅲ和第Ⅷ cbbL 基因簇,说明壤质黏土中可能存在较多的新的自养固碳微生物. 砂质黏壤土 cbbL 基因型主要分布在第 V、第Ⅷ和第Ⅸ cbbL 基因簇,分别占砂黏壤土 cbbL 基 因型 总 数 的 11%、 13% 和 24%,与 Bradyhizobium、Mesorhizobium 和 Nitrobacter 等根瘤菌以及 Mycobacterium sp.、Thermomonospora curvata、Actinobacteria、Nocardia brasiliensis 等放线菌同源.这些结果表明壤质黏土和砂质黏壤土中含细菌 cbbL 基因的兼性自养微生物群落组成差异很大.

3 讨论

自养微生物在农田土壤中广泛存在,可以通过 卡尔文循环固定大气中的 CO₂,对提高农田生态系 统的碳吸收和碳储存有着重要意义^[1,2].本研究对 不同质地稻田土壤进行碳同位素连续标记培养实 验,110 d 培养结束后,两种质地土壤中均检测出了

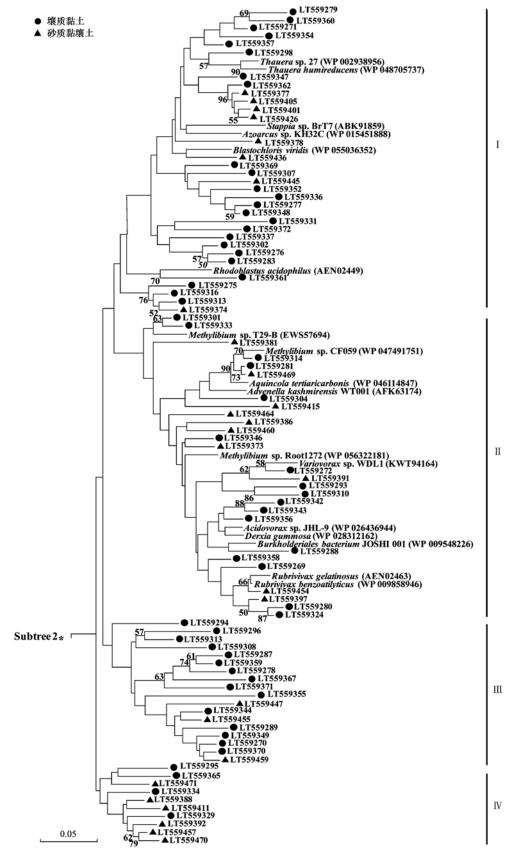
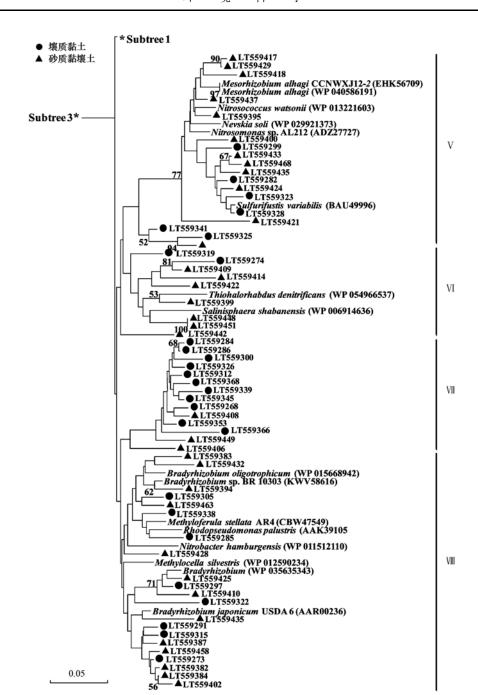


图 2 基于 cbbL 基因 164 个氨基酸序列片段构建的自养固碳微生物系统进化树

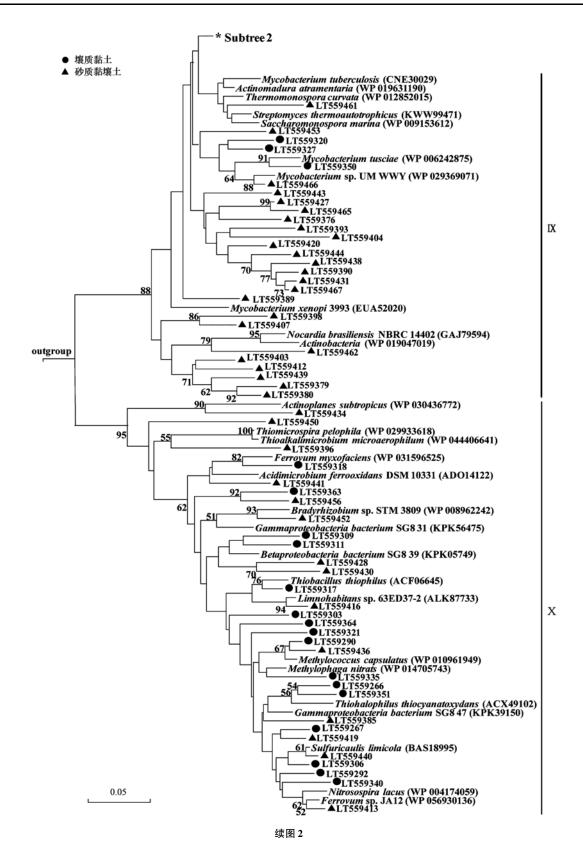
Fig. 2 Phylogenetic tree of cbbL-containing bacteria based on partial cbbL sequences (164 amino acids)



续图2

较高的 ¹℃-SOC含量,其含量介于 95.80 ~ 141.66 mg·kg⁻¹,高于 Yuan 等^[1]报道的稻田土壤自养微生物碳同化量(46.41 ~ 64.61 mg·kg⁻¹),这说明土壤微生物确实发挥了 CO₂ 同化能力.以往的研究结果显示仅在光照培养土壤中检测到 ¹℃标记碳,而在黑暗培养土壤中几乎检测不到 ¹℃^[1],说明在本实验条件(光照培养)下,两种质地土壤中的 ¹℃主要由自养微生物固定.但不同质地土壤 ¹℃含量存在差异,壤质黏土 ¹℃-SOC含量均显著高于砂质黏壤土,表明土壤质地显著影响自养微生物碳同化量,壤质黏土自

养微生物具有更高的固碳潜力,这可能与自养固碳微生物的多样性和群落结构有关.与植物根际沉积碳相似^[21],土壤自养微生物同化碳(新碳)进入土壤后也向土壤微生物量碳库(MBC)和土壤可溶性碳库(DOC)活性碳库转化,但壤质黏土自养微生物同化碳向 MBC 和 DOC 转化的量(¹⁴C-MBC和¹⁴C-DOC)高于砂质黏壤土,这表明土壤质地影响了自养微生物同化碳在土壤中的转化和稳定性.与砂质黏壤土相比,壤质黏土养分含量较高,因而有利于维持更高的土壤微生物数量、代谢活性以及功能多样性,促



进微生物对土壤新碳的转化利用^[2,8],使得壤质黏土¹·C-MBC和¹·C-DOC高于砂质黏壤.

质地不同的土壤其物理化学性质也不同,而土 壤的理化性质又是影响自养固碳微生物数量、活性 和群落结构的重要生态因子^[1,7,9]. 因此,土壤质地差异可能会引起自养固碳微生物群落结构及其多样性的变化,从而最终导致不同质地土壤自养固碳微生物碳同化量的差异. 本研究通过构建 cbbL 基因克

隆文库和系统发育分析发现本研究土壤中自养固碳 微生物以兼性自养菌为主,且土壤质地对自养固碳 微生物群落结构和多样性产生了显著影响,壤质黏 土比砂质黏壤土具有更高的自养固碳微生物多样 性. 这可能与两种质地土壤理化性质不同有关. Xiao 等[9] 通过冗余分析发现土壤自养微生物群落 结构是土壤环境因子综合作用的结果,土壤自养固 碳微生物多样性受土壤黏粒含量、阳离子交换量和 土壤有机质含量的显著影响. 黏粒含量和阳离子交 换量越高,土壤中可溶性营养物质的积累越多,土壤 养分越不容易流失[22,23],越有利于自养固碳微生物 (兼性自养微生物)的生长繁殖和活性的增强[1,7,9]. 本研究中,两种质地土壤环境和养分含量差异较大, 与砂质黏壤土相比,壤质黏土黏粒含量、土壤有机 质、全氮含量和阳离子含量更高. 因此, 壤质黏土 可为土壤兼性自养微生物的生长提供更丰富的基质 和能源物质,维持更高的自养固碳微生物的多样性, 促进兼性自养菌的生长和代谢活性,因而壤质黏土 自养微生物同化碳量较高;而砂质黏壤土黏粒组分 含量低,营养物质和养分积累少,能为兼性自养菌提 供的碳源和能源有限,自养固碳微生物多样性相对 较低,生长代谢活动较弱,因而砂质黏壤土自养微生 物同化碳量较低. 此外,运用 Libshuff 命令分析结果 显示壤质黏土和砂质黏壤土的细菌 cbbL 基因文库 存在显著性差异(P<0.05),说明土壤质地影响了 细菌 cbbL 的群落结构. 壤质黏土 cbbL 基因序列与 Rhodoblastus acidophilus, Blastochloris Thauera humireducens、Mehylibium sp. 和 Variovorax sp. 等具有较高的同源性,而砂质黏壤土 cbbL 基因 序列则与一些根瘤菌和放线菌同源. 稳定同位素核 酸探针(DNA-SIP)分析结果表明, Rhodoblastus acidophilus (原 Rhodopseudomonas)、Mehylibium sp.、 Variovorax sp. 和 Bradyhizobium 等具有较强的 CO, 同化能力,是参与稻田土壤 CO2 固定的主要自养微 生物类群^[24]. 目前对放线菌 CO, 同化能力的研究 较少,尽管许多放线菌全基因组序列中含有 cbbL 基 因,不过,已证实的能通过卡尔文循环固定 CO, 的 放线菌菌群还非常少[25~27]. 因此,不同质地土壤自 养固碳微生物群落组成的差异可能是导致其碳同化 量差异的另一重要原因,因为不同类型固碳细菌同 化 CO₂ 的能力不同.

4 结论

(1) ¹C-CO, 连续标记培养 110 d 后, 壤质黏土

¹⁴C-SOC含量范围为 129.83 ~ 141.66 mg·kg⁻¹,而砂质黏壤土 ¹⁴C-SOC含量范围为 95.80 ~ 112.91 mg·kg⁻¹,壤质黏土 ¹⁴C-SOC平均含量显著高于砂质黏壤土,说明壤质黏土有更高的固碳潜力.

- (2)土壤质地影响了土壤自养微生物同化碳在土壤中的转化、 $^{14}\text{C-CO}_2$ 连续标记培养 110d 后,壤质黏土 $^{14}\text{C-MBC}$ 和壤质黏土 $^{14}\text{C-DOC}$ 平均含量分别为49. 16 mg·kg $^{-1}$ 和8. 10 mg·kg $^{-1}$,显著高于砂质黏壤土 $^{14}\text{C-MBC}$ 和 $^{14}\text{C-DOC}$ 平均含量(33. 21 mg·kg $^{-1}$ 和4. 18 mg·kg $^{-1}$).
- (3)土壤质地改变了土壤自养固碳微生物群落结构(P<0.05)和多样性,壤质黏土具有更高的自养固碳微生物多样性.

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