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抗生素类制药废水厌氧消化产物急性毒性的检测

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摘要:为判断抗生素类制药废水厌氧消化产物对功能菌群自身的毒害作用,采用发光细菌法研究了制药废水中残留抗生素及厌氧消化中间产物的单独及联合毒性(15 min-IC₅₀). 试验表明,厌氧消化中间产物乙醇、乙酸、丙酸和丁酸对发光杆菌的半抑制浓度(15 min-IC₅₀)分别为 19. 40、20. 71、10. 47 和 12. 17 g·L⁻¹,其毒性大小顺序为:丙酸 > 丁酸 > 乙醇 > 乙酸. 不同类抗生素氨苄青霉素、卡那霉素、林可霉素和环丙沙星对发光杆菌的半抑制浓度分别为 3. 99、5. 11、4. 32 和 5. 63 g·L⁻¹,其毒性大小顺序为:氨苄青霉素 > 林可霉素 > 卡那霉素 > 环丙沙星. 厌氧消化中间产物,氨苄青霉素-厌氧消化中间产物,环丙沙星-厌氧消化产物对发光细菌的联合毒性呈相加作用;卡那霉素-厌氧消化中间产物、林可霉素-厌氧消化中间产物、4 类抗生素与厌氧消化中间产物对发光细菌的联合毒性呈协同作用. 研究结果表明,可使用发光细菌评价抗生素类制药废水厌氧消化处理的可行性,并为工程开发提供操作依据.

关键词:发光细菌; 抗生素; 制药废水; 厌氧消化; 毒性

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Acute Toxicity of Antibiotics and Anaerobic Digestion Intermediates in Pharmaceutical Wastewaters

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Abstract: In order to determine the toxicity of antibiotics and anaerobic digestion intermediates on anaerobic treatment of pharmaceutical wastewaters containing antibiotics, the single and joint toxicities of some antibiotics and intermediates to *Photobacterium phosphoreum* were tested by using the 15-min half inhibitory concentration (15 min-IC₅₀) at pH = 7.00 ± 0.05. The results showed that the 15 min-IC₅₀ of ethanol, acetate, propionate and butyrate were 19.40, 20.71, 10.47 and 12.17 g·L⁻¹, respectively, which indicated that the toxicity descended in the order of propionate, butyrate, ethanol and acetate. The 15 min-IC₅₀ of Amoxicillin, Kanamycin, Lincomycin and Ciprofloxacin were 3.99, 5.11, 4.32 and 5.63 g·L⁻¹, respectively, so the toxicity descended in the order of Amoxicillin, Lincomycin, Kanamycin and Ciprofloxacin. Using equal effect mixing method, the joint toxicity of four anaerobic digestion intermediates, the four intermediates together with Amoxicillin, Ciprofloxacin, Kanamycin, Lincomycin individually and all together were investigated, which demonstrated that the first three interactions were additive and the last three were synergistic. The observations have laid a foundation for control and optimization of anaerobic biotechnology for pharmaceutical wastewater containing antibiotics.

Key words: luminescent bacterium; antibiotics; pharmaceutical wastewater; anaerobic digestion; toxicity

厌氧消化技术具有无需供氧、产生沼气、减排污泥等诸多优点,已在环境和能源工程中广泛应用 $^{[1-5]}$. 若以容积有机负荷(organic loading rate, OLR)作为厌氧消化功效指标,现有生产性厌氧消化技术的 OLR(以 COD 计,下同)可达 20 ~ 40 kg·(m³·d) $^{-1}$,实验室厌氧消化技术的 OLR 可达 86 ~ 306 kg·(m³·d) $^{-1}$,厌氧消化技术还有很大的潜力可挖 $^{[6-8]}$.

厌氧消化技术为混菌发酵技术,其对有机物的转化主要由水解产酸菌群、产氢产乙酸菌群、产甲烷菌群协作完成.由于各菌群的生长和代谢性能不同^[9,10],易积累中间产物(包括乙醇与各种挥发性有机酸),对厌氧消化功能菌群具有抑制作用,最终会

破坏厌氧消化过程的高效性与稳定性. 较高的挥发酸浓度不仅对甲烷菌有抑制作用, 对有机物质的降解也有反馈抑制作用^[11],高浓度的 VFA 会抑制产甲烷菌的活性,造成"酸中毒",当总 VFA 浓度超过13 000 mg·L⁻¹时厌氧消化即停止^[12]. 此外,抗生素生产废水是一类难以生物处理的工业废水^[13~15],其中的抗生素对功能菌具有毒害作用. 这些毒物的单独或联合作用均可抑制功能菌群,导致厌氧消化反应器效能的崩溃.

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研究制药废水厌氧处理中抗生素和厌氧消化中间产物的单独及联合毒性,有助于诊断厌氧消化障碍,推进制药废水厌氧消化处理工艺的研发.发光细菌的发光强度对各种毒物毒性的响应具有综合性,与一般监测手段相比,具有成本低、速度快和灵敏度高等优点,发光细菌检测法已广泛应用于环境监测[16]、食品安全性监测[17]等领域的污染物的毒性评价[18-20].但有关制药废水厌氧消化中抗生素与消化中间产物的联合毒性,迄今未见文献报道.因此,本课题采用发光细菌检测法,试验了制药废水中所可能包含的4类抗生素和厌氧消化中间产物的单独及联合急性毒性,以期为抗生素类制药废水的高效厌氧处理提供理论依据和调控参数.

1 材料与方法

1.1 试验菌种

明亮发光杆菌(Photobacterium phosphoreum) T3 变种,购自中国科学院南京土壤研究所. 培养液及培养基配方参照文献[21]. 菌液制备方法如下:将安瓿管中存放的冻干粉溶于经灭菌的 0.5 mL 3% NaCl 溶液中,再迅速转入 50 mL 培养液中,20℃恒温培养,每 24 h 转接 1 次斜面,将培养好的第三代斜面置于 4℃冰箱中,作为菌种备用;菌种接入培养基平板上,20℃恒温培养 16~22 h 后,加入 5 mL 3% NaCl 至平板上,经适当振荡后将菌体洗下,再用漩涡振荡器将菌液混匀,浓度调节至合适范围,作为试验菌剂备用.

1.2 试验仪器

DXY-2 型生物毒性测试仪(中国科学院南京土壤所); YXQ-SG41-280 型手提式压力蒸汽灭菌锅(上海华线医用核子仪器有限公司); ZHWY-Z102C型恒温摇床培养箱(上海智城仪器制造厂); Five easy 型 pH 计(瑞士梅特勒托利多); 微量进样器(10 μL,美国 Agilent 公司).

1.3 试验试剂

无水乙醇、乙酸、丙酸、正丁酸均为分析纯,购 于上海国药集团化学试剂有限公司;氨苄青霉素、 卡那霉素、林可霉素和环丙沙星均购于生工生物工 程(上海)有限公司.

1.4 试验方法

1.4.1 化合物单独毒性测定

在18~22℃下,以3% NaCl 溶液作稀释液,将 乙醇、乙酸、丙酸、丁酸、氨苄青霉素、卡那霉素、 林可霉素和环丙沙星均配成6~7个浓度梯度,用 3% NaCl 溶液配制的 0.1 $\operatorname{mol} \cdot \operatorname{L}^{-1}$ NaOH 与 0.1 $\operatorname{mol} \cdot \operatorname{L}^{-1}$ HCl 调节各溶液 pH 至 6.95 ~ 7.05 之间. 混合均匀后,取 2 mL 待测溶液于发光强度测试管中,每个浓度设置 3 个平行,以 2 mL 3%的 NaCl 作为空白对照. 取 10 pL 试验菌剂于各测试管中,振荡摇匀,15 min 后用生物毒性测试仪测定其发光强度 $\operatorname{E}^{[22]}$. 利用空白对照发光强度的平均值 LU_0 和各浓度 3 组平行样发光强度的平均值 LU_0 计算发光杆菌的相对发光强度 RLU_0 即:

$$RLU = \frac{LU}{LU_0}$$
 (1)

由所得的线性回归方程式求出相对发光强度为50%时所对应的测试化合物的浓度,即为该化合物对发光细菌发光强度的半抑制浓度(15min-IC₅₀).本试验相关统计学分析均采用 Microsoft Excel 2003或者 DPS 8.0 软件.

1.4.2 化合物联合毒性测定

由单独毒性试验所得到的抗生素和厌氧消化中间产物的 15min-IC₅₀,采用等效浓度混合法^[23]进行联合毒性试验,配置 6 组混合物: A (乙醇+乙酸+丙酸+丁酸+氨苄青霉素)、B (乙醇+乙酸+丙酸+丁酸+专那霉素)、C (乙醇+乙酸+丙酸+丁酸+林可霉素)、E (乙醇+乙酸+丙酸+丁酸+环丙沙星)、F(乙醇+乙酸+丙酸+丁酸+氨苄青霉素+卡那霉素+林可霉素+环丙沙星),测定混合物对发光细菌相对发光强度,求得混合物的联合毒性大小,测定方法同1 4 1 节

等效浓度混合(按照单一化合物的 15min-IC₅₀ 之比混合)后,可按式(2)计算各单一化合物混合过程所占的质量浓度比.

$$\chi_{i} = \frac{IC_{50(i)}}{IC_{50(a)} + IC_{50(b)} + \dots + IC_{50(i)} + \dots IC_{50(n)}}$$
(2)

式中 χ_i 为毒物a、b、n 在混合物中所占的质量浓度比, $\sum \chi_i = 1$; $IC_{50(i)}$ 为毒物a、b、n 在单一体系中的 IC_{50} .

对多元混合体系,可采用以下计算法判定混合体系的预期半抑制浓度:

$$\frac{1}{{\rm IC}_{50(E)}} = \sum \frac{\chi_i}{{\rm IC}_{50(i)}} \quad (i = a, b, \cdots, n) \quad (3)$$

式中, $IC_{50(E)}$ 为多元混合物预期的 IC_{50} .

由单一化合物的 $15 \min - IC_{50}$ 和式(3),可计算出 多元混合物的 $15 \min - IC_{50(E)}$,可按式(4)计算 R 值.

$$R = \frac{IC_{50(E)}}{IC'_{50}} \tag{4}$$

式中,IC₅₀为实测的混合物对发光细菌发光强度的 半数抑制浓度.

联合作用的类型可按 R 值判定: 若 R < 0.4,联合作用则判为拮抗作用; 若 R > 2.5,联合作用则判为协同作用; 若 0.4 < R < 2.5,联合作用则判为相加作用.

2 结果与讨论

2.1 化合物的单独毒性

为了解厌氧消化中间产物及4类抗生素的单独毒性大小,以明亮发光杆菌为靶,分别测定了4种厌氧消化中间产物与4类抗生素的急性毒性,试验结果见图1~2. 从中可知,7种化合物对发光细菌的相对发光强度与其浓度呈良好的相关性. 由线性

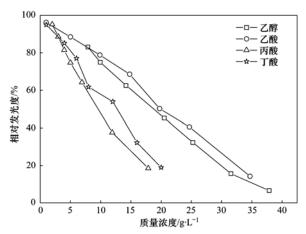


图 1 厌氧消化中间产物对明亮发光杆菌的浓度-反应关系

Fig. 1 Relationship between the concentration of anaerobic digestion intermediates and the RLU of *P. phosphoreum*

拟合所得方程(见表 1)可得化合物的单一半抑制浓度(15min-IC₅₀). 乙醇、乙酸、丙酸与丁酸的15min-IC₅₀值分别为19.40、20.71、10.47 和12.17 g·L⁻¹, 其毒性的相对大小为丙酸(1.98) > 正丁酸(1.70) > 乙醇(1.07) > 乙酸(1.00). 氨苄青霉素、卡那霉素、林可霉素与环丙沙星的15min-IC₅₀值分别为3.99、5.11、4.32 和5.63 g·L⁻¹,4 类抗生素单独毒性的相对大小为氨苄青霉素(1.41) > 林可霉素(1.30) > 卡那霉素(1.10) > 环丙沙星(1.00).

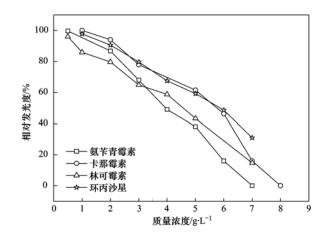


图 2 4 类抗生素对明亮发光杆菌的浓度-反应关系

Fig. 2 Relationship between the concentration of antibiotics and the RLU of *P. phosphoreum*

厌氧消化中间产物对厌氧消化功能微生物的急性毒性的检测方法鲜见报道. Blum 等 $[^{24}]$ 研究发现,有机毒物对产甲烷菌的 IC_{50} 值与其对发光细菌的 IC_{50} 值之间有良好的线性关系($R^2=0.9056$),表明用发光细菌检测法评价厌氧消化中间产物对消化功能菌群的毒性是可行的.

表 1 化合物的单独急性毒性1)

Table 1 Single toxicity of chemicals to P. phosphoreum

化合物	线性回归方程	相关系数	IC ₅₀ /g·L ⁻¹	化合物	线性回归方程	相关系数	IC ₅₀ /g·L ⁻¹
乙醇	Y = -2.601X + 100.45	0. 990 3	19. 40	氨苄青霉素	Y = -15.849X + 113.31	0. 988 5	3. 99
乙酸	Y = -2.477X + 101.28	0. 993 6	20.71	卡那霉素	Y = -14.143X + 121.15	0. 957 3	5. 11
丙酸	Y = -4.876X + 101.15	0. 982 4	10. 47	林可霉素	Y = -11.975X + 101.78	0. 988 0	4. 32
丁酸	Y = -4.080X + 99.64	0. 989 1	12. 16	环丙沙星	Y = -10.891X + 111.33	0. 988 6	5. 63

1)Y 为相对发光度(%),X 为化合物浓度(g·L⁻¹),下同

乙醇对细菌的毒害机制:一是进入蛋白质分子,使蛋白质变性沉淀^[25];二是溶解细胞膜上的脂质而增大细胞膜的通透性^[26];三是抑制细菌酶活,阻碍正常代谢^[27].挥发性有机酸(volatile fatty acid, VFA)可瓦解细胞膜内外质子梯度而阻碍 ATP 合成^[28,29].3种 VFA 的单独毒性与其 pK_a 值有关.在

水溶液中,一元酸(HA)存在如下平衡:

$$HA = H^{+} + A^{-}$$

$$K_{a} = \frac{[H^{+}][A^{-}]}{[HA]}$$

$$\frac{[HA]}{[HA]_{0}} = \frac{[HA]}{[HA]_{0} + [A^{-}]} =$$

$$\frac{1}{1 + K_{a} / [H^{+}]} = \frac{1}{1 + 10^{pH - pK_{a}}}$$

20℃ 时, 乙酸、丙酸和丁酸的 pK_a 值分别为 4.75、4.87、4.81^[30]. pH 为 7.0 时,对于浓度同为 5 000 $mg \cdot L^{-1}$ 的乙酸、丙酸和丁酸,其未解离态浓度分别为 27、38 和 33 $mg \cdot L^{-1}$,以丙酸的毒性最大. 厌氧发酵液 pH = 7.0 时,浓度为5 000 $mg \cdot L^{-1}$ 的丙酸盐对甲烷产率的抑制为 22% ~ 38% [23],与发光细菌检测法所得的结果基本一致.

4 种抗生素的分子结构如图 3 所示. 氨苄青霉素属青霉素类抗生素,其作用机制较复杂,其结构与位于肽聚糖亚单位上的 D-丙氨酰-D-丙氨酸结构相似,可抑制转肽反应酶,从而阻断肽聚糖的合成而导

致渗透裂解,它只作用于处在生长中的细菌;也可激发一种称做细菌 holin 的特殊蛋白,从而在原生质膜上形成孔洞或损伤,导致膜泄漏而死亡[31].卡那霉素属氨基糖苷类抗生素,结构中含环己烷和氨基糖,可结合到核糖体亚基上而干扰蛋白质合成,也可直接抑制蛋白质合成,还可引起 mRNA 遗传信息的错读.林可霉素属大环内酯类抗生素,可与 50S 核糖体亚基的 23S rRNA 结合,抑制蛋白质合成过程中肽链的延长[32].环丙沙星属喹诺酮类抗生素,可结合于 DNA 促旋酶复合物上,抑制细菌 DNA 促旋酶或 DNA 拓扑异物酶 II 发挥作用,阻碍 DNA 复制、转录、分裂期细菌染色体的分离以及其他涉及 DNA 的细胞过程[31].

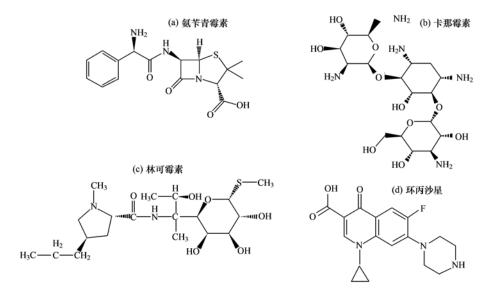


图 3 氨苄青霉素、卡那霉素、林可霉素及环丙沙星结构式

Fig. 3 Chemical structure of amoxicillin, kanamycin, lincomycin and ciprofloxacin

2.2 化合物的联合毒性

为了了解厌氧消化中间产物的联合毒性以及其与不同类抗生素的联合毒性,以明亮发光杆菌为靶,采取

等效应混合法,分别测定了多元混合物的急性毒性.

厌氧消化中间产物以及抗生素与消化中间产物 对明亮发光杆菌的联合毒性结果见图 4. 从中可知,

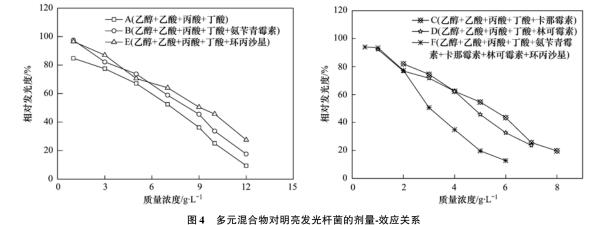


Fig. 4 Relationship between the concentration of chemical mixtures and the RLU of P. phosphoreum

多元混合物联合作用下对发光细菌的相对发光强度 与其浓度具有良好的相关性.对图 4 中的试验数据 进行线性拟合,所得的回归方程见表 2.

经计算,多元混合物组合 A、B、C、D、E、F 的 R 值分别为 0.779、1.176、2.630、2.860、1.539、3.047,因此可判定厌氧消化中间产物(4元混合

物)、消化中间产物-氨苄青霉素(5元混合物)、消化中间产物-环丙沙星(5元混合物)的联合毒性均呈相加作用;消化中间产物-卡那霉素(5元混合物)、消化中间产物-林可霉素(5元混合物)、消化中间产物-4类抗生素(8元混合物)的联合毒性均呈协同作用.

表 2 多元混合物的联合毒性

Table 2 Joint toxicity of chemical mixtures to P. phosphoreum

	, i	1		
多元混合物	线性回归方程	相关系数	实测 IC ₅₀ /g⋅L ⁻¹	预期 IC ₅₀ /g·L ⁻¹
A(乙醇+乙酸+丙酸+丁酸)	Y = -2.586X + 102.09	0. 990 2	20. 141	15. 686
B(乙醇+乙酸+丙酸+丁酸+氨苄青霉素)	Y = -4.758X + 104.01	0. 985 5	11. 351	13. 348
C(乙醇+乙酸+丙酸+丁酸+卡那霉素)	Y = -10.833X + 105.90	0. 987 4	5. 160	13. 572
D(乙醇+乙酸+丙酸+丁酸+林可霉素)	Y = -11.446X + 103.70	0. 987 2	4. 691	13.414
E(乙醇+乙酸+丙酸+丁酸+环丙沙星)	Y = -6.066X + 103.89	0. 988 7	8. 884	13. 676
F(乙醇+乙酸+丙酸+丁酸+氨苄青霉素+ 卡那霉素+林可霉素+环丙沙星)	Y = -16.428X + 105.13	0. 978 0	3. 356	10. 225

从毒理学上看,毒物结构相近、性质相似、靶器官相同或作用机制相同时,联合效应呈相加作用;而当毒物毒害的途径、方式和作用部位各不相同时,所产生的联合效应为独立作用;当某一毒物能够促进细胞对其他毒物的吸收和积累或阻碍其降解和排泄时,则这些毒物的联合效应为协同作用^[23].

消化中间产物(4元混合物)的联合毒性,因结构相似且作用机制相同而呈相加作用. 厌氧消化中间产物-氨苄青霉素(5元混合物)的联合毒性也呈现相加作用,究其原因,可能是其作用的靶器官类同且作用机制接近. 尽管厌氧消化中间产物-环丙沙星(5元混合物)中各化合物对微生物的作用机制不同,但厌氧消化中间产物的存在不会促进细胞对环丙沙星的吸收和积累从而阻碍 DNA 复制和修复,因此其联合毒性也呈现相加作用. 厌氧消化中间产物-卡那霉素(5元混合物)、厌氧消化中间产物-林可霉素(5元混合物)、厌氧消化中间产物-4种抗生素(8元混合物)的联合毒性均因靶器官不同且作用机制互异,且可相互促进细胞对其他化合物的吸收从而呈协同作用.

3 结论

(1) 厌氧消化中间产物乙醇、乙酸、丙酸和丁酸对发光杆菌发光强度的半抑制浓度(15 \min -IC₅₀)分别为 19. 40、20. 71、10. 47 和 12. 17 g·L⁻¹, 其毒性大小顺序为: 丙酸 > 丁酸 > 乙醇 > 乙酸. 四类抗生素氨苄青霉素、卡那霉素、林可霉素、环丙沙星对发光杆菌发光强度的半抑制浓度(15 \min -IC₅₀)分

别为 3. 99、5. 11、4. 32 和 5. 63 g·L⁻¹, 其毒性大小顺序为: 氨苄青霉素 > 林可霉素 > 卡那霉素 > 环丙沙星.

(2) 厌氧消化中间产物(4元混合物)、厌氧消化中间产物-氨苄青霉素(5元混合物)、厌氧消化中间产物-环丙沙星(5元混合物)的联合毒性 R 值分别为 0.779、1.176、1.539,对发光细菌的联合毒性呈相加作用. 厌氧消化中间产物-卡那霉素(5元混合物)、厌氧消化中间产物-林可霉素(5元混合物)、厌氧消化中间产物-4类抗生素(8元混合物)的联合毒性 R 值分别为 2.630、2.860 与 3.047,其对发光细菌发光强度的联合毒性呈协同作用.

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